## Gold Nanoparticle: Synthesis, Functionalization, Enhancement, Drug Delivery and Therapy: A Review

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ASTRACT The excellent properties copyrighted by a numb physical, chemical and colloidal or suspended optical properties. The effective role in improvin nanoparticles of many treatment. Despite the di in the field of drug deliver a treatment and nano ca vaccine and genetics, so	and advantages of golden nanoparticles are er of publisher roles. There are different biological methods possible in preparing nanoparticles with pharmacodynamic and surface of golden nanoparticles plays an ng performance and efficacy as a carrier of drugs, especially in the field of cancer fficult challenges faced by gold nanoparticles y, these molecules are a great opportunity as rrier to deliver drugs to anticancer, antibiotic, a comprehensive study must be conducted	to find out all the pharmacokinetics an long periods. <b>Keywords:</b> Gold nanoparticles AuN nanoparticles <b>Correspondence:</b> Ihsan A. Mohammed Ministry of Health and Environmer Babylon, Iraq E-mail: <u>ehsan2013.mohammed@gmail</u> <b>DOI:</b> <u>10.31838/srp.2020.6.127</u> @Advanced Scient	d cytotoxic properties of cells for Ps, pharmacological, inorganic t, Babylon Health Directorate, .com tific Research. All rights reserved

#### INTRODUCTION

The goal of pharmaceutical dosages is to enhance the dynamic capacity of treatment and are protected by the rights to be written and preserved. Therefore, this kinetic capacity of the drug must be increased using nanoparticles and is a molecule of size less than 100 nanometers [1-4]. In 1857 Faraday and others first prepared colloidal gold which is a solution or suspension of nanoparticles of gold nanoparticles in a liquid, most of which is water by reducing gold chloride with phosphorous and produces an intense red or colloidal color of blue / purple gold [5-9].

During the 20 th century, procedures like TEM (transmission electron microscopy) and AFM (atomic force microscopy) allowed the precise imaging of nanoparticles. nanoparticles AuNPs(GNPs) are inorganic Gold nanoparticles consist of a gold atom inner core and negative groups on the surface [10-15]. The surface can be simply functionalized for active targeting by adding a monolayer of surface ligands. In addition, they can be modified by different chemical and physical methods, AuNPs (GNPs) for biomedical uses are mainly synthesized using the colloidal preparation method using a gold metal precursor, a reductant, and a stabilizer [16,17]. This methodology permits accurate control of the optical and electrical properties that powerfully depend on the size scale from 1nm to 100 nm and various shapes as nanosphere, nanocage, nanoshell, and nanorod of the produced AuNPs(GNPs) [18-22]. GNP's innovative drug delivery systems have many benefits over other conventional as well as to nanocarrier drug delivery systems. Some of these advantages are enumerated here; (a) GNPs have characteristic optical, physical, and chemical properties due to their different size and shape. (b) GNPs provides high drug loading capacity due to high surface area (c) GNPs are readily available for biofunctionalization with biomolecules such as proteins, enzymes, carboxylic acid and DNA and are biocompatible; (d) GNPs have controlled uniform dispersity; (e) Due to nano-scale and uniform dispersion of GNPs they can reach to the targeted site of action with

blood flow easily, They are safe and noncytotoxic to the normal cells and (f) GNPs are easily synthesized by different methods of synthesis. Challenges of GNP as a nanocarrier drug delivery system are (a) Factors affecting biodistribution, pharmacokinetics, and pharmacological properties need to be clarified. (b) Long term cytotoxicity effects must be studied. (c) Eliminate the inflammatory and some polymer coatings immune response triggered by (d) Economics [23-30]. It is of crucial need to identify the exact GNP's physicochemical properties that permit optimum gastrointestinal absorption and accumulation in the site of action after oral intake and these properties are greatly determined by surface charge and size of GNPs. In the study intra-oesophageal administration of radiolabelled positive surface charges, GNPs of 2.8 nm size and negative surface charge GNPs with different sizes range 1.4 to 200 nm into healthy adult female rats. After one day and by gamma spectroscopy the amount of the GNPs in organs and tissues was determined [31-35].

Individual designs have been developed based on the nature of the tissues and human cells targeted in the treatment because the size of the nanoparticles has determined the highest values of accumulation and the load granted by the drugs. The highest accumulation of secondary organs in the human body occurred in the size of the AuNPs (GNPs) at 1.4 nm that charged with a more positive charge Of the negative charge, compared with 18 nm particles, which showed the highest accumulating values in the heart and brain cells [36-40].

#### GOLD NANOPARTICLES

Gold particles are ranged from large size particles to small nanparticles, they are differ in their physical properties like color, size, state of matter and activity. As well as gold nanoparticles differ from each other by shape and cluster to give a lot of characteristic shapes. The shape of gold nanoparticles affect their optical properties therefore, triangular nanoparticles have attractive optical properties in contrast to other shapes (Figure 1) [41].



Figure 1: Various shapes of gold nanoparticles

Using of plant extract is important to purify gold nanoparticles and determine their medical applications. They are therapeuticall used to target anticancer medication to tumor cells to kill them by hyperthermal treatment as well as used for biomolecular ultrasensitive detection. Florescent nanoparticles applied to make imaging of certain enzymes and metabolite inside cancer cells .gold nanorods have a lot of applications in in the field of biosensing, photo thermal therapy and gene delivery because of their absorptive properties in the visible and near infrared region [42-46].

Gold nanoparticles highly used in CT imaging as molecular probes rather than iodine due to its higher absorption coefficient attributed to higher atomic number and electron density [47,48]. The biological safety and large surface area of gold nanoparticles are other beneficial properties of them to be used in biomedical purposes. Colloidal nanoparticles are very small in size like that of DNA and proteins and easily prepared, therefore highly used to target cells and tissues due to easy of the entrance to inside these cells. Gold nanoparticles can be used as vaccine carriers and mainly epidermal DNA vaccine delivery by gene gun gold nanoparticles due to safety and binding to various organic molecules. by coating nanoparticles with temperature labile polymer, they can be used as drug carriers [49-55].

## SYNTHESIS STRATEGIES

A number of methods have been used in chemical, physical, and biological methods in synthesis and stability strategies for golden nanoparticles GNPs.

## 3.1. The chemical reduction method

The preparation of GNPs by the chemical reduction method consist of two steps: (a) reduction step by use agents like formaldehyde, hydroxylamine, sugars, carbon monoxide, hydrogen peroxide, sulphites, acetylene, hydrogen; citric and oxalic acids (b) stabilization step by using agents like sulphur ligands, phosphorus ligands, polymers and surfactants. To avoid the aggregation of the GNP [56-60].

#### 3.2. The Green methods

Green chemistry synthesis routes are environment friendly and non-toxic. A facile green biosynthesis method for the preparation of gold nanoparticles of size 25 + 7 nm was reported by using natural biomaterial egg shell membrane (ESM) [61-69]. In this method ESM was immersed in aqueous solution of HAuCl4 without using any reductant. Another green synthetic approach was developed to synthesized gold sononanoparticles of size 5 - 17 nm by using high-power ultrasounds and sodium dehydrate. Gold nanoparticles were successfully synthesized by adopting sun light irradiation method and were modified with folic acid and capped by 6-mercaptopurine. In this method solar energy was used to reduce the gold salt. A new green chemistry method for the preparation of gold nanoparticles has been reported, in which gold nanoparticles were formed in aqueous NaCl solution from the bulk gold substrate by natural chitosan without using any external stabilizer and reductant [70]. Gold nanoparticles of size 15 - 80 nm are also synthesized via another green synthetic route. In this method HAuC14 was reduced by using citrus fruits juice extracts [Citrus limon, Citrus reticulate and Citrus sinensis]. Edible mushroom was also used for the synthesis of gold nanoparticles via sunlight exposure [71,72].

## 3.3. The Citrate reduction (Turkevich method)

The most common method for synthesis gold nanoparticles AuNPs is the Turkevich reductive method, although there are many other methods. The Turkevich method was discovered in 1951, and some modifications were made to the methods of its construction by the scientist Frens' group, where this process of formation includes reducing citrate with gold particles to produce nanoparticles with a size of 20 nanometers. Particles with sizes ranging from 16 to 147 nanometers can be produced by adjusting the proportions of the ester components with gold. The mechanics of this method were examined by Peng's group in 2007(Schematic 1) [73-81].



Schematic 1: shown of Two Pathways for the formulation of gold nanocrystals by citrate reduction [82]

The nucleation process followed in the synthesis of nanoparticles AuNPs can be constructed in two ways, either by reducing citrate or by determining pH values. Any of these methods can be accomplished in three Pathways, smoothing the nanowires to dots, random attachment to polycrystalline nanowires, and naming nucleation. pH values may give the most common nucleation shift pathway information [83-85].

#### 3.4. The Brust-Schiffrin method

The Brust – Schiffirin reaction pathway involves building thermally stable, air-stable nanoparticles AuNPs with reduced dispersion values. This method was discovered by the scientist Brust–Schiffirin and his group in 1994, through which the size of nanoparticles with a range of 1.5 nm to 2.5

can be controlled. In this way, two reaction pathways were used to obtain an effective surface reaction during the development and growth process. AuCl<sup>-4</sup> has been transformed from its aqueous solution to another organic solvent, which is staining with the aid of a phase transfer reagent, followed by a reductive pathway with sodium borohydride and dodecanethiol that shown in scheme 2. In comparison with the method of reducing the citrate, this method includes the formation of a hydrophobic mineral group and then its dissolution without a change in the properties of the formed particles. The method of Brust-Schiffirin has become more extensive when using percaptophenol which provides greater stability than other pathways [86-89].



Scheme 2: Shows the Reaction for Brust–Schifrin method [82]



Fig. 2: Shows of the proposed processes contributory in ligand Replacement reaction on Au-MPCs[82].

The gold nanoparticles have become more stable by a proposed set of functional lecandes. Several modifications have occurred in a practiced method, and Murray's group has discovered protection for gold particles by monolayer-protected gold clusters (MPCs) as a multifunctional chemical reactor that has been widely used and gives

encouraging results. The alkanethiol ligands (RS) have been used through Replacement reactions (Figure 2) [90-93].

#### 3.5. Polymer-based synthesis of GNP

Size and shape of GNP consider a vital part in colloidal gold preparation. The interaction of polymers with GNP greatly affect the size diversity and stability of particles. Study overcome this condition by using antitumor drug entrained into a PEG capping layer with an acid-labile spacer. According to this stated records, such a system will considerably increase the efficiency of both release and intracellular uptake of cytotoxic drug [94-98].

#### 3.6. The Physical method

#### 3.6.1. The Electrochemical method

The GNPs were prepared electrochemically using a simple two-electrode cell, with reduction of the cathode and oxidation of the anode. The electrochemical construction of nanoparticles was first considered by Reetz, in 1994. This procedure has been considered to be superior to other techniques of nanoparticle formation, due to its lower processing temperature, low cost, high quality, modest equipment and ease of process managing [99-102].

#### 3.6.2. The Seeding growth method

According to this method of preparation, GNP of diameters 5-40 nm with a narrow dispersity in size were prepared. Particle size can be well ordered by change ratio of seed to metal salt. This method has the gain of being an easy, quick, and low cost; whereas the used of trisodium citrate as a source of OH ions in the seeding step and sodium borohydrate (NaBH<sub>4</sub>) as a reductant [103-106].

## 3.6.3. Ultraviolet-induced photochemical synthesis of GNPs

The potential chemistry attributed to the use of GNPs in magnetic devices, photocatalysis, fabrication and aerosol is powerfully influence by controlling morphology and dimension features of the prepared nanoparticle. As many researches have stated, the photoreduction process permits the preparation of single crystallite GNPs. The preparation of GNPs with controllable size was effectively achieved by photochemistry [107-110].

## 3.6.4. Ultrasound aided synthesis of GNP

Generator of ultrasound wave was used for a water bath with controlled temperature for the ultrasonic-aided reduction of gold precursor in presence of 2-propanol. Various stabilizers have been used during this preparation method, such as citrate, disulphide and several dendrimers, for reproducibility and tunability reasons [111-114].

#### 3.6.5. Laser ablation synthesis of GNP

Accurate and reproducible outcomes have been obtained by laser ablation procedure, in terms of shape and size attributes. So, the pulsed laser process which needs simultaneous evaporation and condensation occurrences for gold represents a comprehensive physical method that can be effectively apply to yield GNPs with tuneable properties. The preparation needs reduction of HAuCl<sub>4</sub> by laser beam of a 532 nm wavelength, producing GNPs with 5 nm and lower in size. In this method, solution of sodium dodecyl sulphate (SDS) has been consider as a model and study the effect of both laser and concentrations on the size and shape of the prepared GNPs. GNPs got by this method

useful in immunochromatographic assay labelling [115-118].

### 3.7. The Biological method

By this method GNP are prepared by microorganisms, enzymes, and plants.

#### 3.8. The Microbial synthesis of GNP

The necessity of eco -friendly and low cost preparation of GNPs by utilize microorganisms are highlighted due to no dangerous by-products. The process has been assumed that enzymes like ligninases, laccases and reductases are used in nucleation and growth of GNPs. Numerous factors affect the preparation and stability of GNPs like substrate concentration, pH, temperature, and static condition. Nevertheless, there are many works on enhancing these procedures. Some state that Klebsiella pneumonia mediated synthesis of GNPs and synergetic effect of antimicrobial activity to many bacterial pathogen S. aureus, E. coli, S. Epidermidis and P. aeruginosa. Other colloidal gold mediated synthesis is soil isolation of fungus Penicillium crustosum used in success whole preparation of GNPs mediated by extracellular proteins [119-123].

#### 3.9. Plant mediated synthesis of colloidal gold

Lately, the use of plants for the preparation of GNP reflect area of concern, because of their low cost, availability, nontoxic nature and eco-friendliness. In latest years, the synthesis of GNPs using different plants such as, Aloe vera, Cinnamomum camphora, Azadirachta indica and Coriandrum sativum have been stated [124-126].

## FUNCTIONALIZATION OF GNPs

Surface biofunctionalization is one of the most encouraging features of GNPs in the biomedical field and can be modified with different biomolecules, such as antibodies, peptides, and DNA. There are two types of functionalization interactions. One of them is noncovalent interactions, the other is covalent interactions. Noncovalent alterations take place through hydrophobic entrapment, electrostatic interactions, and van der Walls forces [127-130]. The binding is not solid enough to produce stable surfaces tolerate the required incubation conditions and washing steps, particularly in biological researches. Thus, the impact of the surrounding medium's pH and ionic strength is important to consider. In contrast, covalent modifications, which make use of linker molecules or immediate chemical attachment, gives more reproducibility and stability. Covalent interaction is able to tolerate a high salt concentration and is very stable under thermal settings. However, covalent interaction is more complicated, from time to time needing concentrated preparation effort on the ligands. The simplicity of this surface modification by noncovalent and covalent modifications can be used in biodiagnostic and biosensing and for specific biological targeting [131-134].

#### THE BOOST OF GNPs

The drug delivery function of GNPs can be enhanced in two ways, one way by prolongation of their plasma half-life and therefore accumulation within cells and the other way by increasing cellular uptake and drug release inside targeted cells. polyethylene glycol conjugation with GNPs will decrease their clearance by macrophages and reticuloendothelial system (RES) [135-138]. Because it provides a steric barrier against macrophages as well as to potentiate penetration and retention process. Zwitterions could be used also for the same purpose because it is neutral and therefore, prevent interaction with macromolecules of cells like receptors and proteins. Conjugation with ligands like folate, peptides or antibodies will provide better targeting of GNPs to cancerous tissues by binding to specific receptors on diseased cells [139-142].

Size is also an important factor that determines the half-life of GNPs since large particles are more rapidly cleared by macrophages and RES, although the ideal size for cellular uptake till now it is unknown. Wong et al. founded an inverse relationship between size and cellular uptake in contrast to Yue et al. who founded that larger size GNPs have higher cellular uptake than smaller ones. Kumar et al. concluded the same findings as with Wong et al [143-145].

Morphology of GNPs is another critical factor that affects the cellular uptake, studies found that spherical GNPs have a higher rate and extent of cellular uptake than rod-shaped ones. Xie et al. found that triangular GNPs have higher cellular uptake than star and rods shaped ones. Surface charge of GNPs is another important factor in addition to size and shape that determine the rate of cellular uptake, highly positive GNPs are more efficient in cellular uptake than negatively charged one since cells are negatively charged. Studies found that bovine serum albumin and installed glucose could be used as ligands on GNPs surface to enhance binding to specific tissue receptors and thus increase uptake. pH and temperature are critical factors that affect drug release in diseased tissues so acidic pH around cancer cells determine the release of drug by using PH responsive formulations and heat-sensitive materials that

control drug release. Another example is by using glutathione responsive materials with GNPs to enhance the release of drugs inside cancer cells since they are rich in glutathione [146-148].

#### APPLICATIONS OF NANOPARTICLES

The gold nanoparticles GNPs (AuNPs) have unique properties due to their electrical and magnetic advantages. A large number of studies concerned with studying AuNPs molecules in biomarking, chemical sensing, human biological medicine, electronic phototherapy, thermal phototherapy, nanotechnology medical imaging, DNA diagnosis of both types, treatment transfer techniques, study of transfer and attribution of reagents to AuNPs particles in the treatment of cancers (Table 1) [149-151]. Various sensors based on the action of the reagents assigned to the AuNP<sub>s</sub> particles based on the principle of the color change of the detector can identify and estimate the various metal ions. The detection paths were used by the sensor method to determine the ions of lead, mercury, copper and zinc in the water. Inert elements such as gold play an active role in the work of the sensors, due to the ratio of the total area of the gold particles to their size. Biomolecules assigned to the nanoparticles have been used in the work of medical biomedical devices such as MPA (Mercaptopropionic Acid) assigned to AuNPs. The linear range of the sensors ranges from 0.25 mM to 1.25 mM at a concentration of 0.025 mM glucose [152] (Figure 4). The work of LSPR biosensors is based on the surface plasmon resonance of the gold nanoparticles, and catalixar derivatives have been used for the purpose of enhancing the efficiency of the work with the gold nanoscale sensors. DNA has been detected using peptides in the nanosensors and has proven highly efficient in identifying the amino acids and quaternary ammonium ions and prenium as well. The anandium tin oxide electrode used with the TiO<sub>2</sub> compound based on AuNPS particles was used to estimate the catechol (CC) and hydroquinone (HQ) with the help of volumetric pathways. Catechol was estimated in tea extract using this method [153-155].

Shape	Size	Application	
Nano rod	2-5 nm	Drug delivery and photothermal therapy [15].	
Hollow particle	25 nm	Photo-electronics, catalysis and cancer therapy [12]	
Triangular particle	3.85-7.13 nm	Highly effective against <i>E. coli</i> and <i>K</i> . pneumonia [23]	
Faceted particle	50-100 nm	Effective, reproducible, and stable large area substrates for NIR SERS [near infra-red surface enhanced Raman spectroscopy] [25]	
Nanocube	50 nm	Field enhancement applications and refractive-index sensing [28]	
Nanocage	50 nm	Effective molecular contrast agent for nonlinear endomicroscopy imaging [19] and in vivo medical applications [29]	
Nanobelt	Thickness, ~80 nm, Width, ~ 20 μm, Length, ~0.15 m.	Strain sensors	
Branched particle	90 nm	Substrates for SERS-based imaging of kidney cells [30]	

Table 1: Shapes of	gold nano	particles and	their applications
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#### 6.1. Surface plasmon resonance (SPR)

The optical properties of gold nanostructures are influenced by surface plasmon resonance (SPR). Plasmon resonance is the electronic frequency that occurs on the surface of gold nanoparticles when its frequency coincides with that of the magnetic wave. Therefore, the area of nanoparticles plays an effective role in the effect of plasmon resonance(SPR) [156-158].

#### 6.2. Sensor material

AuNPs have photovoltaic properties that make them very suitable for chemical sensing, and the inactivity properties of these particles make them biomedical materials of very low toxicity compared to some biocompatible technological polymers [159-162].

#### 6.3. Nucleotides sensor for homogeneous detection

In 1997, the discovery of the ability of AuNPs to modify the surface of nanoparticles using threatened nucleotides, its study of stability and hydrobiological degree. This discovery paves the way for finding highly sensitive and selective materials that make building nanoparticles of gold more stable, cooperative and highly corrosive in stimulating surface interaction [163-165]. Figure 3 shows the result of a

reaction of a mixture of DNA-AuNPs to form a polymeric network, accompanied by changes in color for this reaction from red to purple. This change is governed by differences in the size of the assembly and the optical properties of the particles. Also, the intense color of the particles of AuNPs is due to SPRs and hence the gold nanoparticles appear Formed with a size of 13 nm and a red color. Surface plasmon show relatively narrow absorption values at the wavelength of 520 nm, with an ultraviolet spectrum, while some AuNPs show a purple color at a range of absorption values of 520 to 574 nm. This means a red displacement in the AuNPs [166-168]. The greatest importance lies in discovering the DNA to collect the AuNPs as a lining on the DNA, and this way to discover the DNA is much better than the traditional methods based on fluoridation, but there are some bad advantages from using color strategies in detection because they have a specific detection limit, especially when using concentrations. The picomular was reduced to the concentration of the nanomolar due to the limited sensitivity of this process, but low detection limits can be avoided and modified when using low-enriched nucleotides by dithiane epidrosterone functionalized oligonucleotides [169-172].



Fig. 3: shows the Properties of DNA-functionalized AuNPs [82].

6.4. Nucleotides sensor for heterogeneous detection High sensitivity techniques have been developed to detect DNA and RNA strings in the same in the same test sample. This codification has been used by Letsinger and his group by a sandwich containing fixed capture. Through the high concentrations of the target sequence, the antibody antibody attached to the surface of the nanoparticles can be observed with the naked eye, and if low concentrations are used this will enhance the silver shells that will increase the size of the AuNPs to the micrometer and thus facilitate its detection process. The Letsinger method is 100 times more sensitive than traditional Fluorescence methods [173-175]. Human RNA samples have been used to detect mRNA matrices, and with the development of other tests, new methods have been worked out that are more sensitive to measurement. AuNPs of different sizes can be used at the same time to detect the target sequence with a dual- color reading(Figure 4) [176-178].



Fig 4: shows of scanometric of DNA detection assay [82].

#### 6.5. Protein sensor

The color change from red to violet resulting from dispersion can be considered as well as the use of AuNPs aggregation in color sensors to detect DNA and RNA. It can also hybridize DNA. A number of studies have demonstrated the use of lectin-sugar in sensing sugar when an interaction between sugars and lectin occurs and that this reaction is appropriate for biological sensing. Russells and his group studies have shown that the stable AuNPs of mannose sugar can be used to detect lectin (Figure 5) [179-182]. The interaction between sugar monomers, which are weak and thus the sensitivity of the sensor, decreases when using a polymerase chain reaction chain reaction (RAFT) reaction pathways because preparation in this way provides functional groups ending with the triol group, which can correspond to the surface area of the gold particles to make AuNPs. Scheme 3 illustrates the stages of constructing polyacrylamide associated with mannose sugar using the RAFT polymerization method. Reducing the thiocarbon functional group to a reducing group is thiol, through which mannose sugar is combined with the gold surface that contributes as stabilizers to prevent the aggregation of AuNPs. Genetically modified bacteria introduced into the

polymer have the ability to form strong bonds with protein because of the many bonds that are affected with protein. The RAFT technique was used to polymerize the modified acrylamide with glucose sugar which showed a clear association with the surface of AuNPs (Scheme 4) [183-185]. The complexes prepared to detect the blood components did not show any interaction between the red blood cells and thus show their ability in biological biological applications. The highest importance is shown in the use of glucosamine prepared in protein sensors (Scheme 5) [186]. Regardless of the studies, glucose-polymer binding with Concanavalin A was demonstrated by a multicomponent bonding. Sugars can be used as terminal modifiers instead of as peripheral monomers (Scheme 6) [187]. The linked lectin, which is assigned to the AuNPs particles, was used to detect the sugars. Once the lectin was linked with the glucose sugar, it would change the color of the glucose to red due to the assembly process and thus the detection speed and stability of the AuNPs particles with the glucose could be adjusted and a polymeric coating was obtained to reach the optimal detection state [188-190].



Fig. 5: (a) shows that the mannose stabilized with AuNPs (b) Changes in UV-Vis spectra for the mannose stabilized with AuNPs



Scheme 3: shows the Synthesis of the glycopolymer and the preparation of the polymer-immobilized AuNPs



Scheme 4: shows the Synthesis of poly(B-D-1-glucopyranosyl hydroxyethyl acrylamide) [82].



Scheme 5: Shows the Synthesis of glucosamine functionalized polymethacrylate



Scheme 6: Shows the Synthesis of polymer-stabilized AuNPs [82].

#### 6.6. Common drugs for cancer Therapy

Constructive pathways for clinical treatment of cancer are confined. Surgical restriction, chemotherapy and irradiation are some common tactics used for cancer therapy but these approaches are not only toxic, non-specific but also cause various side effects [191-193]. Cancer patients undergoing radio and chemotherapy face drug resistance such as cisplatin, cancer related fatigue (CRF) and several cardio vascular effects such as cardiomyopathy, ischemia, arrhythmias, hyper tension, thromboembolism, pericardial diseases or heart attack as they damage the cancer cells along with the destruction of healthy cells [194-196].

Besides of some conventional approaches, chemotherapy remains the primary treatment for cancer but it is not much restorative because of various side effects caused by unspecified drug distribution in the body due to several chemotherapeutic agents such as cytotoxic drugs [197-199]. Paclitaxel (PTX) shows cytotoxicity against different types of cancer so it is considered to be an essential chemotherapeutic drug with limited therapeutic effects due to toxicity caused by poor water solubility and selectivity. Because of these shortcomings, effective approaches should be considered.

Chemotherapeutic agents cause various side effects in cancer patients such as nephrotoxicity, vomiting, myelosuppression, severe nausea, ototoxicity and neurotoxicity due to CDDP (cisplatin) administration whereas gastro intestinal disturbances, acute nausea, vomiting, stomatitis, alopecia baldness, neurologic disturbances, bone marrow, aplasia, cumulative cardio toxicity and bone marrow depressant effects due to doxorubicin administration [202,204].

Due to fluoropyrimidines (5FU), methotrexate, irinotecan and cisplatin patient may suffer from adverse diarrhea and constipation [205]. When treated with temsirolimus as a single drug anemia, hyperglycemia, stomatitis, hypophosphatemia, interstitial lung disease and pneumonia were reported in patients suffering from advanced renal cell carcinoma [206]. Each year, large numbers of deaths are caused by cancer because lack of selectivity, drug targeting ability, inefficient metastatic tumor therapy and drug resistant tumor cells (Table 2). Therefore advanced chemotherapic treatments have been needed to kill cancerous cells [207]. Today in order to avoid side effects fuscous of scientists are shifted towards natural products which still needs to prove their effectiveness [208].

Table 2: Conventional intravenously adr	ministered drugs and their side effects
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Chemotherapeutic agent	Cancer type	Possible short-term side effect
Carboplatin (Paraplatin)	Cancers of the ovary, head and neck, and lungs	Decrease in blood cell counts, hair loss (reversible), confusion, nausea, vomiting, and/ordiarrhea
Cisplatin (Platinol, Platinol-AQ)	Cancers of the bladder, ovary, and testicles	Decrease in blood cell counts, allergic reaction, including a rash and/or labored breathing, nausea and vomiting that usually occurs for 24 hours or longer, ringing in ears and hearing loss, fluctuations in blood electrolytes, and kidney damage
Doxorubicin (Adriamycin)	Breast cancer, lymphoma, and multiplemyeloma	Decrease in blood cell counts, mouth ulcers, hair loss (reversible), nausea and vomiting, and heart damage
Paclitaxel (Taxol)	Cancers of the breast, ovary, and lung	Decrease in blood cell counts, allergic reaction, nausea and vomiting, loss of appetite, change in taste, thin or brittle hair, joint pain (short term), and numbness or tingling in fingers or toes
Fluorouracil (5-FU)	Cancers of the colon, breast, stomach, and head and neck	Decrease in blood cell counts, diarrhea, mouth ulcers, photosensitivity, and dry skin

6.7. GNP conjugated Antibiotics Drug Delivery System In vitro evaluation of GNPs conjugated antibiotics shows significant reduction in minimum inhibitory concentration for streptomycin and kanamycin conjugated GNPs and slight decrement in GNPs conjugated ampicillin minimum inhibitory concentration value compared to its free drug form. In addition to improvement in heat stability for all studied antibiotics values in their GNPs conjugated form, this formulation provides a modern, safe, and effective strategy for the treatment of bacterial infections with GNPs conjugated drug delivery syste [209-212]. New approach utilizing bacterial toxins to specifically deliver antimicrobials drugs to the sites of infections and activation of smart drug release from GNPs. Coupling of chitosan functionalized GNPs to the liposomes surface lead to increase liposomes stability and prevent immature drug release in physiological environments or labeled storage condition. Nevertheless, once bacterial the stabilized liposomes toxin secreted these protected liposomes find bacteria, the toxins will penetrate liposome membranes and generate pores, through which the antibiotic drug released. The released antibiotic subsequently exerts its antimicrobial effects on the toxin-secreting bacteria as a smart drug delivery system [213,216]. In this study vancomycin used as a model bactericidal dug to methicillin-resistant staphylococcus aureus (anti-MRSA) by using GNP-Liposomes nanocarriers, result shows that the encapsulated

vancomycin prepared as GNP-liposomes can entirely release within 24 h in response MRSA bacteria toxin and inhibit MRSA growth. Other work study GNP conjugated antibiotic (ampicillin, streptomycin and kanamycin) as drug candidate for this drug delivery system. Where bactericidal efficacy evaluated in different bacterial strains Staphylococcus aureus, Escherichia coli and Micrococcus luteus and find there is more significant reduction in minimal inhibitory concentration with greater bactericidal activity. In addition, all these GNPs conjugated antibiotics revealed higher storage condition stability (heat and UV) in comparison to their free forms [217-219].

#### 6.8. GNP and gene delivery systems

Gold nanoparticles have been used as an alternative delivery system for many drugs, proteins, RNAs (siRNAs), plasmid DNAs (pDNAs), peptide and chemotherapy. Surface modification systems for functional genes assigned to GNPs, such as adding thiol, amine and carboxyl groups, have been modified. The modification can be made by adding positive polymers. The GNP-RNA nanoscale system has been widely used in siRNA connection. The siRNA synthesis was changed by adding a thiol group to improve the efficacy of GNPs. The modified siRNA synthesis with thiol showed strong adsorption on the surface of GNPS particles within human cells, directing the SERNA group directly and the cause of RNA interactions with the reductive cell cytoplasm unlike the direct association with the surface of  $GNP_s$  particles. Binding of siRNA ends to chemical bonds with  $GNP_s$  particles attached to the polymers. This binding by reducing bonds leads to the formation of a fissionable environment and the formation of smart siRNA in the cytoplasm in the cell. This work depends on the control factor of pH values [220-222].

The induction of target protein synthesis was initiated by the plasmid DNA delivery vectors (pDNAs), whether the delivery systems used are viral or non-viral, these conductive systems are characterized by high efficiency in the transmission of infection and this is the main reason for their repeated uses, despite the limited efficiency of the nonviral delivery system However, it can be a substitute for viral delivery systems due to the possibility of modifying the design of non-viral delivery systems, in addition to that, the small size of GNPs particles are highly efficient in transporting very high payloads for each individual GNP carrier due to this The largest surface area of a particle to a ratio of volume [223-225].

#### 6.9. GNPs Recent Advances in Vaccines

In 1928, the scientist Cole and his group discovered the ability of the immune system to respond to cancerous tumors, as stated in his study that vaccination with stimulating cancer cells can increase the degree of immune response against tumors, thus finding vaccines against cancers. The high selectivity of these vaccines makes them useful as a treatment for the destruction of cancerous tumor cells. Mankind is very acceptable. Studies concerned with the localization of antigens associated with cancerous tumors (TAAS) and tumor-specific antigens (TSAS), and efforts have been combined to develop approaches for these studies. It includes an immunization method against tumors by nature of tumor cells and genetically engineered tumor cells engineering [226-228].

All vaccination studies aim at working with vaccination strategies and improving immune response and developing them to be able to destroy cancer cells only without other natural cells without damage. Gold nanoparticles have proven their ability to visualize the delivery of effective antigens to cancer cells and have helped stimulate the work of T-lymphocyte during effective chemotherapy. Nanoparticles of 15 to 50 nm are very effective in delivering these antigens against more than 45 pathogens of parasite, bacterial and viral infections [229-231].

Various forms of nanoparticles of gold and of different sizes have been used as an antigen holder in oncology treatments in general by pairing with the targeted physico-chemical antigen. Extensive studies were conducted on the role of the SH group in protein molecules and their association with gold nanoparticles. GNPs are also coupled with a multisugar-bond or protein link before being antigen-loaded. The latest engineering designs have been used to make GNPs suitable for loading vaccines and antigens by improving the shape, size and surface characteristics. These modern designs have also contributed to making nanoparticles suitable for improving immunity and proliferation in the lymph nodes and activating T-cell response to the antigen [232-234]. 6.10. Gold nanoparticles for improved and enhanced phototherapy.

The therapeutic techniques of gold nanoparticles are divided into four main sections that work as an anti-cancer treatment, which is PTT photodynamic and dynamic radiotherapy. The main function of these particles is dynamic photodynamic therapy PDT and photothermal therapy PTT. The PDT technology is non-invasive and works using light-sensitive agents (Optical Optimizer) as this technique works to produce reactive oxygen in different types (ROS) for the purpose of damaging target cancer cells, as it uses hydrophobic photocatalysts such as phthalocyanin and porphyrins, and thus it needs a PEGylated nanocomputer for the purpose of improving complete dissolution in water, and the PDT provides an effective protection system (5 -fluorouracil) when coupled with .GNPs, the GNPs provide an effective carrier and carrier for treatment to divorce the payload on the ultraviolet with a maximum wavelength. The function of PTT is a preventive measure to destroy cancer cells at high temperatures in GNPs when irradiated by light. GNPs have the ability to absorb SPR if irradiated with light using short laser waves [235-238].

PTT was coupled with GNPs for the purpose of creating a lethal pulse of cancerous cells. Using PTT and PDT with X-rays only for a specific area of cancer, and therefore the activity of using X-rays is more effective than in the infrared in the activation of PTT and PDT. When using the gold element in irradiation, which has a higher absorption coefficient and less toxicity, therefore, the killing of prostate cancer cells has been improved by about 15% - 20% when assigned to GNPs by proton radiation [239-241].

The targeting systems of GNPs particles differ in the degree of delivery of each drug according to the effects and the size and quantity of materials associated with the GNPs particles. For example, when the GNPs are in small capsules, they are sensitive to heat, especially when LSPR is used in the process of operation when medicines are released from the capsules. After destroying the nanostructures of GNPs [242-244].

# 6.11. Applications of targeted delivery systems from cells to clinics

Nanoscale platforms are used to diagnose different types of cancer. The target delivery system for nanoparticles acts as anti-cancer platforms such as audited liposomes, tails nanoparticles and albumin based drug carriers. Nanoparticles have been used to diagnose cancers by photogrammetry during surgery and bilateral imaging. In general, nanoscopy has become effective in early diagnosis of cancerous and human diseases. Dual contrast factors allow the diagnosis of areas of interest to be examined in two independent ways, namely MRI detectors and the use of fluorine agents. In the case of MRI, there is very high sensitivity and accuracy in diagnosis. Fluoroscopy compensates for MRI in the case of overcoming restrictions during the surgical procedure being imaging Uni-style. This study offers a wide application in the surgery of cancerous tumors in the ovaries in the early stages of the tumor, with specific imaging of the tumor for patients suffering from ovarian cancer through folic acid receptors. Gold nanoparticle platforms are a versatile platform that provides desirable results for diagnostic systems as they are prepared in a size range from 1.5 to 10 nanometers. Usually gold nanoparticles are made using hydrogen tetrachloride and the treatment is loaded by covalent bonding. Gold nanoparticles have particularly beneficial properties in diagnosing cancers, compressible heart size, monobifurcation, low toxicity, light scattering properties, ease of manufacture, and the ability to bind to biomolecules via Au-S. Gold nanoparticles have demonstrated the potential of targeted heat therapy in conjunction with a chemotherapy agent in treating MDR tumors [245-247].

Heo et al. Described. (2012) golden nanoparticles as a diagnostic platform where they operate on the surface using biotin, PEG, rhodamine Bletaed beta-cyclodextrin (beta-CD) and baclitaxel. Paklitaxel is a compound that is included with beta-CD and has then been coupled with golden nanoparticles. Laboratory studies indicate that golden nanoparticles have a high tendency toward cancer cells such as MG63, A549 and HeLa compared to NIH<sub>3</sub>T<sub>3</sub> cells. Golden nanoparticles have demonstrated a toxic effect on HeLa cancer cells [248-250].

Effective and smart gold nanoparticles have been developed where DOX is coupled with nanoparticles via Au-S using peptide tape bases, Cys-Pro-Leu-Gly-Leu-Alu-Ala-Ala-Gly-Gly (CPLGLAGG) where the cleavage is Specifically by protease (Figure 4) and demonstrated by animal experiments after injection of functional gold nanoparticles in tumor-bearing mice, excessive protease [251,253].

## ADVANTAGES OF GOLD NANOPARTICLES

Gold nanoparticles mediated drug delivery systems have many advantages over other nanocarriers as well as to conventional drugs. Gold nanoparticles have been widely used as acancer antigen and in tumor therapies [254]. Some advantages are listed here; (i) Gold nanoparticles have unique optical [255], physical and chemical properties [256] due to their size and shape [257]; (ii) Gold nanoparticles have high surface area [258] which provide dense drug loading; (iii) These particles are biocompatible [259] and are readily available for conjugation with small biomolecules such as proteins, enzymes, carboxylic acid, DNA, and amino acids [260]; (iv) Gold nanoparticles have controlled dispersity [261]; (v) Due to small size and uniform dispersion they can easily reach to the targeted site with blood flow [262]; (vi) They are non-cytotoxic to the normal cells [263]; and (vii) Gold nanoparticles are easily synthesized by various methods [264].

Good characteristics in the using gold nanoparticles as a drug carry and delivery system

- ➤ Control the ratio of the volume to the surface area of the gold nanoparticles easily to achieve the goal of their use.
- ➤Control the release and preservation of the drug during transport and storage, and achieve increased effectiveness and disposal of side effects.

- ➤Achieving the highest capacity of the particles to load and integrate the drug without any chemical reaction.
- ► Ease of attaining the targeting sites with nanoparticle surface targeting.
- The delivery of medications can be achieved with high success to the mouth, eye, nose, intravenous injection, etc.
- ➤Stability and reliability in drug delivery can be enhanced by avoiding combination of drugs and nanoparticles.
- Restricting the motion of the drug particles that are supported by the surface of the AuNPs reduces drug drain.
- The surface of the particles can be easily modified if they are in a solid / liquid phase.

## DISADVANTAGES OF NANOPARTICLES

#### 8.1. Potential toxicity

While the small size of nanoparticle is what makes them so useful in medicine, it is also the factor that might make them potentially dangerous to human health [265].

#### 8.2. Environmental concerns

Artificially manufacture nanoparticles will be new to the environment in type and quantity and would constitute a new class of non biodegradable pollutants [266].

#### IDEAL PROPERTIES OF NANOPARTICLES

- ≻It can be used as a natural or synthetic polymer
- ≻cheap
- ≻Toxic free
- ≻Easily decomposes
- ≻It does not clot easily
- ► It does not affect the immune system
- $\succ \mbox{The diameter of nanoparticles must be less than 100 nanometers$
- Platelet aggregation does not occur during the assignment of drugs to nanoparticles in the treatment of hematology.
- > The use of nanoparticles does not cause tissue infections

## CONCLUSION

Gold nanoparticles have revolutionized the field of medicine because of its widespread applications in targeted drug delivery, imaging, diagnosis, and therapeutics due to their extremely small size, high surface area, stability, noncytotoxicity, and tunable optical, physical and chemical properties. Functionalized gold nanoparticles with various biomolecules such as proteins, DNA, amino acids, and carboxylic acids have been used to provide excellent cancer therapy and provide excellent drug delivery systems. Targeted delivery and programmed release of therapeutic drugs to the specific site is achieved by using gold nanoparticles because they can bear high drug load and release it to the specific site through various administration routes and can interact with the cancerous cells. Side effects of conventional drugs have been minimized by conjugation with gold nanoparticles and they increase the quality life of patients.

### THE DISAGREEMENT OF INTEREST

The authors declare that there are no conflicts of interest.

#### REFERENCES

- 1. Cuiping Yao, Luwei Zhang, Jing Wang, Yulu He, Jing Xin, Sijia Wang, Hao Xu, and Zhenxi Zhang, Gold Nanoparticle Mediated Phototherapy for CancerJournal of Nanomaterials, 19 Dec 2016.
- Lian-Hua Fu, jun yang, Jiefang Zhu, Ming-Guo Ma, Synthesis of Gold Nanoparticles and Their Applications in Drug Delivery, Springer, Cham., November 2017, pp 155-191.
- R. V. Nair, H. Santhakumar, and R. S. Jayasree, "Gold nanorods decorated with a cancer drug for multimodal imaging and therapy," Faraday Discuss. 207, 423–435 (2018).
- Link S, El-Sayed MA. (2002). Shape and size dependence of radiative, non-radiative and photothermal properties of gold nanocrystals. International Reviews in Physical Chemistry, 19 (3), 409-453.
- McCoy CP, Rooney C, Edwards CR, Jones DS, Gorman SP. (2007). Light-triggered molecule-scale drug dosing devices. Journal of the American Chemical Society, 129(31), 9572–9573.
- 6. D. Liu and D. T. Auguste, "Cancer targeted therapeutics: from molecules to drug delivery vehicles," Journal of Controlled Release, vol. 219, pp. 632–643, 2015.
- G. Chirico, P. Pallavicini, and M. Borzenkov, "Physical properties of gold nanostars," in Gold Nanostars, pp. 25–42, Springer, Berlin, Germany, 2015.
- M. A. Zaimy, N. Saffarzadeh, A. Mohammadi, H. Pourghadamyari, P. Izadi, A. Sarli, L. K. Moghaddam, S. R. Paschepari, H. Azizi, S. Torkamandi, and J. Tavakkoly-Bazzaz, "New methods in the diagnosis of cancer and gene therapy of cancer based on nanoparticles," Cancer Gene Ther. 24(6), 233–243 (2017).
- J. P. M. Almeida, A. Y. Lin, E. R. Figueroa, A. E. Foster, and R. A. Drezek, "In vivo gold nanoparticle delivery of peptide vaccine induces anti-tumor immune response in prophylactic and therapeutic tumor models," Small 11(12), 1453–1459 (2015).
- R. P. Brinās, A. Sundgren, P. Sahoo, S. Morey, K. Rittenhouse-Olson, G. E. Wilding, W. Deng, and J. J. Barchi, Jr., "Design and synthesis of multifunctional gold nanoparticles bearing tumor-associated glycopeptide antigens as potential cancer vaccines," Bioconjug. Chem. 23(8), 1513–1523 (2012).
- A. L. Parry, N. A. Clemson, J. Ellis, S. S. R. Bernhard, B. G. Davis, and N. R. Cameron, "'Multicopy multivalent' glycopolymer-stabilized gold nanoparticles as potential synthetic cancer vaccines," J. Am. Chem. Soc. 135(25), 9362–9365 (2013).
- 12. A. Pradhan, M. Bepari, P. Maity, S. S. Roy, S. Roy, and S. M. Choudhury, "Gold nanoparticles from indole-3-

carbinol exhibit cytotoxic, genotoxic and antineoplastic effects through the induction of apoptosis," RSC Advances 6(61), 56435–56449 (2016).

- R. Robinson, W. Gerlach, and H. Ghandehari, "Comparative effect of gold nanorods and nanocages for prostate tumor hyperthermia," Journal of Controlled Release, vol. 220, pp. 245–252, 2015.
- J. R. Melamed, R. S. Edelstein, and E. S. Day, "Elucidating the fundamental mechanisms of cell death triggered by photothermal therapy," ACS Nano, vol. 9, no. 1, pp. 6–11, 2015.
- H. Wang, J. Han, W. Lu, J. Zhang, J. Li, and L. Jiang, "Facile preparation of gold nanocages and hollow gold nanospheres via solvent thermal treatment and their surface plasmon resonance and photothermal properties," Journal of Colloid and Interface Science, vol. 440, pp. 236–244, 2015.
- J. Huang, M. Guo, H. T. Ke et al., "Rational design and synthesis gamma Fe2O3@Au magnetic gold nanoflowers for efficient cancer theranostics," Advanced Materials, vol. 27, no. 34, pp. 5049–5056, 2015.
- C. Iodice, A. Cervadoro, A. L. Palange et al., "Enhancing photothermal cancer therapy by clustering gold nanoparticles into spherical polymeric nanoconstructs," Optics and Lasers in Engineering, vol. 76, pp. 74–81, 2016.
- Z. Zhang, J. Wang, X. Nie, T. Wen, Y. Ji, X. Wu, Y. Zhao, and C. Chen, "Near infrared laser-induced targeted cancer therapy using thermoresponsive polymer encapsulated gold nanorods," J. Am. Chem. Soc. 136(20), 7317–7326 (2014).
- W. P. Savarimuthu, P. Gananathan, A. P. Rao, E.Manickam, and G. Singaravelu, "Protoporphyrin IX-gold nanoparticle conjugates for targeted photodynamic therapy—an in-vitro study," Journal of Nanoscience and Nanotechnology, vol. 15, no. 8, pp. 5577–5584, 2015.
- 20. J. D. Meyers, Y. Cheng, A.-M. Broome et al., "Peptidetargeted gold nanoparticles for photodynamic therapy of brain cancer," Particle & Particle Systems Characterization, vol. 32, no. 4, pp. 448–457, 2015.
- M. Yu, F. Guo, J. Wang, F. Tan, and N. Li, "Photosensitizerloaded pH-responsive hollow gold nanospheres for single lightinduced photothermal/photodynamic therapy," ACS Applied Materials and Interfaces, vol. 7, no. 32, pp. 17592– 17597, 2015.
- C. H. Wang, Y. H. Jiang, X. H. Li, and L. K. Hu, "Thioglucosebound gold nanoparticles increase the radiosensitivity of a triple-negative breast cancer cell line (MDA-MB-231)," Breast Cancer, vol. 22, no. 4, pp. 413–420, 2015.
- 23. P. Li, Y.-W. Shi, B.-X. Li et al., "Photo-thermal effect enhances the efficiency of radiotherapy using Arg-Gly-Asp peptidesconjugated gold nanorods that target  $\alpha v \beta 3$  inmelanoma cancer cells," Journal of Nanobiotechnology, vol. 13, article 52, 2015.
- 24. H. J. Kwon, Y. Byeon, H. N. Jeon, S. H. Cho, H. D. Han, and B. C. Shin, "Gold cluster-labeled thermosensitive

liposmes enhance triggered drug release in the tumor microenvironment by a photothermal effect," Journal of Controlled Release, vol. 216, pp. 132–139, 2015.

- R. Sierpe, E. Lang, P. Jara et al., "Gold nanoparticles interacting with β-cyclodextrin-phenylethylamine inclusion complex: a ternary system for photothermal drug Release," ACS Applied Materials and Interfaces, vol. 7, no. 28, pp. 15177–15181, 2015.
- E. P'erez-Herrero and A. Fern'andez-Medarde, "Advanced targeted therapies in cancer: drug nanocarriers, the future of chemotherapy," European Journal of Pharmaceutics and Biopharmaceutics, vol. 93, pp. 52–79, 2015.
- S. C. Coelho, G. M. Almeida, M. C. Pereira, F. Santos-Silva, and M. A. N. Coelho, "Functionalized gold nanoparticles improve afatinib delivery into cancer cells," Expert Opinion on Drug Delivery, vol. 13, no. 1, pp. 133–141, 2016.
- N. Zhang, H. Chen, A.-Y. Liu et al., "Gold conjugatebased liposomes with hybrid cluster bomb structure for liver cancer therapy," Biomaterials, vol. 74, pp. 280–291, 2016.
- X. Xu, W. Ho, X. Zhang, N. Bertrand, and O. Farokhzad, "Cancer nanomedicine: from targeted delivery to combination therapy," Trends in Molecular Medicine, vol. 21, no. 4, pp. 223–232, 2015.
- A.Wicki, D.Witzigmann, V. Balasubramanian, and J. Huwyler, "Nanomedicine in cancer therapy: challenges, opportunities, and clinical applications," Journal of Controlled Release, vol. 200, pp. 138–157, 2015.
- N. Kotagiri, G. P. Sudlow, W. J. Akers, and S. Achilefu, "Breaking the depth dependency of phototherapy with Cerenkov radiation and low-radiance-responsive nanophotosensitizers," Nature Nanotechnology, vol. 10, no. 4, pp. 370–379, 2015.
- M. Szwed, D. Wrona, K. D. Kania, A. Koceva-Chyla, and A. Marczak, "Doxorubicin-transferrin conjugate triggers prooxidative disorders in solid tumor cells," Toxicology in Vitro, vol. 31, pp. 60–71, 2016.
- N. Gao, H. Sun, K. Dong, J. Ren, and X. Qu, "Goldnanoparticle-based multifunctional amyloid-β inhibitor against Alzheimer's disease," Chemistry 21(2), 829–835 (2015).
- D. A. Gonzalez-Carter, Z. Y. Ong, C. M. McGilvery, I. E. Dunlop, D. T. Dexter, and A. E. Porter, "L-DOPA functionalized, multi-branched gold nanoparticles as brain-targeted nano-vehicles," Nanomedicine (Lond.) 15(1), 1–11 (2019).
- A. Plan Sangnier, R. Aufaure, L. Motte, C. Wilhelm, E. Guenin, and Y. Lalatonne, "Hybrid Au@alendronate nanoparticles as dual chemo-photothermal agent for combined cancer treatment," Beilstein J. Nanotechnol. 9, 2947–2952 (2018).
- L. A. Dykman and N. G. Khlebtsov, Gold Nanoparticles in Biomedical Applications (CRC Press, 2017).
- K. Sztandera, M. Gorzkiewicz, and B. Klajnert-Maculewicz, "Gold nanoparticles in cancer treatment," Mol. Pharm. 16(1), 1–23 (2019).

- L. A. Dykman and N. G. Khlebtsov, "Methods for chemical synthesis of colloidal gold," Russ. Chem. Rev. 88(3), 229–247 (2019).
- C. Yao, L. Zhang, J. Wang, Y. He, J. Xin, S. Wang, H. Xu, and Z. Zhang, "Gold nanoparticle mediated phototherapy for cancer," J. Nanomater. 2016, 1 (2016).
- 40. H. Norouzi, K. Khoshgard, and F. Akbarzadeh, "In vitro outlook of gold nanoparticles in photo-thermal therapy: a literature review," Lasers Med. Sci. 33(4), 917–926 (2018).
- J. B. Vines, J.-H. Yoon, N.-E. Ryu, D.-J. Lim, and H. Park, "Gold nanoparticles for photothermal cancer therapy," Front Chem. 7, 167 (2019).
- AK Khan, R Rashid, G Murtaza and A Zahra, Gold Nanoparticles: Synthesis and Applications in Drug Delivery, Tropical Journal of Pharmaceutical Research July 2014; 13 (7): 1169-1177.
- S. Labala, P. K. Mandapalli, A. Kurumaddali, and V. V. K. Venuganti, "Layer-by-layer polymer coated gold nanoparticles for topical delivery of imatinib mesylate to treat melanoma," Mol. Pharm. 12(3), 878–888 (2015).
- 44. C. S. Kumar, A. Mahesh, M. G. Antoniraj, S. Vaidevi, and K. Ruckmani, "Ultrafast synthesis of stabilized gold nanoparticles using aqueous fruit extract of Limonia acidissima L. and conjugated epirubicin: targeted drug delivery for treatment of breast cancer," RSC Advances 6(32), 26874–26882 (2016).
- N. Rizk, N. Christoforou, and S. Lee, "Optimization of anti-cancer drugs and a targeting molecule on multifunctional gold nanoparticles," Nanotechnology 27(18), 185704 (2016).
- Deb S, Patra HK, Lahiri P, Dasgupta AK, Chakrabarti K, Chaudhuri U. Multistability in platelets and their response to gold nanoparticles. Nanomed: Nanotechnol Biol Med 2011; 7: 376-384. 12.
- Ganeshkumar M, Sastry TP, Sathish Kumar M, Dinesh MG, Kannappan S, Suguna L. Sun light mediated synthesis of gold nanoparticles as carrier for 6- mercaptopurine: Preparation, characterization and toxicity studies in zebrafish embryo model. Mater Res Bull 2012; 47: 2113-2119.
- M. U. Farooq, V. Novosad, E. A. Rozhkova, H. Wali, A. Ali, A. A. Fateh, P. B. Neogi, A. Neogi, and Z. Wang, "Gold nanoparticles-enabled efficient dual delivery of anticancer therapeutics to HeLa cells," Sci. Rep. 8(1), 2907 (2018).
- S. Iram, M. Zahera, S. Khan, I. Khan, A. Syed, A. A. Ansary, F. Ameen, O. H. M. Shair, and M. S. Khan, "Gold nanoconjugates reinforce the potency of conjugated cisplatin and doxorubicin," Colloids Surf. B Biointerfaces 160, 254–264 (2017).
- R. A. Morshed, M. E. Muroski, Q. Dai, M. L. Wegscheid, B. Auffinger, D. Yu, Y. Han, L. Zhang, M. Wu, Y. Cheng, and M. S. Lesniak, "Cell-penetrating peptide-modified gold nanoparticles for the delivery of doxorubicin to brain metastatic breast cancer," Mol. Pharm. 13(6), 1843–1854 (2016).

- M. R. K. Ali, Y. Wu, D. Ghosh, B. H. Do, K. Chen, M. R. Dawson, N. Fang, T. A. Sulchek, and M. A. El-Sayed, "Nuclear membrane-targeted gold nanoparticles inhibit cancer cell migration and invasion," ACS Nano 11(4), 3716–3726 (2017).
- L. Papaioannou, A. Angelopoulou, S. Hatziantoniou, M. Papadimitriou, P. Apostolou, I. Papasotiriou, and K. Avgoustakis, "Folic acid-functionalized gold nanorods for controlled paclitaxel delivery: in vitro evaluation and cell studies," AAPS PharmSciTech 20(1), 13 (2018).
- 53. M. Moghiseh, C. Lowe, J. G. Lewis, D. Kumar, A. Butler, N. Anderson, and A. Raja, "Spectral photoncounting molecular imaging for quantification of monoclonal antibody-conjugated gold nanoparticles targeted to lymphoma and breast cancer: an in vitro study," Contrast Media Mol. Imaging 2018, 2136840 (2018).
- K. Kalimuthu, B.-C. Lubin, A. Bazylevich, G. Gellerman, O. Shpilberg, G. Luboshits, and M. A. Firer, "Gold nanoparticles stabilize peptide-drug-conjugates for sustained targeted drug delivery to cancer cells," J. Nanobiotechnology 16(1), 34 (2018).
- N. Saadat, F. Liu, B. Haynes, P. Nangia-Makker, X. Bao, J. Li, L. A. Polin, S. Gupta, G. Mao, and M. P. Shekhar, "Nano-targeted delivery of RAD6/translesion synthesis inhibitor for triple negative breast cancer therapy," Mol. Cancer Ther. 17(12), 2586–2597 (2018).
- Castro, H.P.S.; Wender, H.; Alencar, M.A.R.C.; Teixeira, S.R.; Dupont, J.; Hickmann, J.M. Third-order nonlinear optical response of colloidal gold nanoparticles prepared by sputtering deposition. J. Appl. Phys.2013, 114, 183104.
- S. M. Pradeepa, S. M. Vidya, S. Mutalik, K. Udaya Bhat, P. Huilgol, and K. Avadhani, "Preparation of gold nanoparticles by novel bacterial exopolysaccharide for antibiotic delivery," Life Sci. 153, 171–179 (2016).
- S. Kalita, R. Kandimalla, K. K. Sharma, A. C. Kataki, M. Deka, and J. Kotoky, "Amoxicillin functionalized gold nanoparticles reverts MRSA resistance," Mater. Sci. Eng. C 61, 720–727 (2016).
- 59. H. R. Ali, M. R. Ali, Y. Wu, S. A. Selim, H. F. Abdelaal, E. A. Nasr, and M. A. El-Sayed, "Gold nanorods as drug delivery vehicles for rifampicin greatly improve the efficacy of combating Mycobacterium tuberculosis with good biocompatibility with the host cells," Bioconjug. Chem. 27(10), 2486–2492 (2016).
- L. Du, S. Suo, H. Zhang, H. Jia, K. J. Liu, X. J. Zhang, and Y. Liu, "The alternative strategy for designing covalent drugs through kinetic effects of pi-stacking on the self-assembled nanoparticles: a model study with antibiotics," Nanotechnology 27(44), 445101 (2016).
- A. Amini, M. Kamali, B. Amini, and A. Najafi, "Enhanced antibacterial activity of imipenemimmobilized on surface of spherical and rod gold nanoparticles," J. Phys. D 52(6), 065401 (2019).

- 62. R. Singh, S. Patil, N. Singh, and S. Gupta, "Dual functionality nanobioconjugates targeting intracellular bacteria in cancer cells with enhanced antimicrobial activity," Sci. Rep. 7(1), 5792 (2017).
- K. O. Shittu, M. T. Bankole, A. S. Abdulkareem, O. K. Abubakre, and A. U. Ubaka, "Application of gold nanoparticles for improved drug efficiency," Adv. Nat. Sci: Nanosci. Nanotechnol. 8(3), 035014 (2017).
- 64. Kim HJ, Matsuda H, Zhou H, Honma I. (2006). Ultrasound- triggered smart drug release from a poly(dimethylsiloxane)- mesoporous silica composite. Advanced Materials, 18(23), 3083–3088.
- L.-H. Peng, Y.-F. Huang, C.-Z. Zhang, J. Niu, Y. Chen, Y. Chu, Z.-H. Jiang, J.-Q. Gao, and Z.-W. Mao, "Integration of antimicrobial peptides with gold nanoparticles as unique non-viral vectors for gene delivery to mesenchymal stem cells with antibacterial activity," Biomaterials 103, 137–149 (2016).
- R. Chowdhury, H. Ilyas, A. Ghosh, H. Ali, A. Ghorai, A. Midya, N. R. Jana, S. Das, and A. Bhunia, "Multivalent gold nanoparticle-peptide conjugates for targeting intracellular bacterial infections," Nanoscale 9(37), 14074–14093 (2017).
- B. Casciaro, M. Moros, S. Rivera-Fernández, A. Bellelli, J. M. de la Fuente, and M. L. Mangoni, "Goldnanoparticles coated with the antimicrobial peptide esculentin-1a(1-21)NH2 as a reliable strategy for antipseudomonal drugs," Acta Biomater. 47, 170–181 (2017).
- L. T. Arayan, H. B. Kim, A. W. Bernardo Reyes, N. T. Xuan Huy, I. H. Hong, K. Lee, J.-H. Yeom, Y. Park, and S. Kim, "The immunomodulatory effect of antimicrobial peptide HPA3P restricts Brucella abortus 544 infection in BALB/c mice," Vet. Microbiol. 225, 17–24 (2018).
- Y. E. Hur, S. Kim, J.-H. Kim, S.-H. Cha, M.-J. Choi, S. Cho, and Y. Park, "One-step functionalization of gold and silver nanoparticles by ampicillin," Mater. Lett. 108., 185–190 (2014).
- X. Li, M. Takashima, E. Yuba, A. Harada, and K. Kono, "PEGylated PAMAM dendrimer-doxorubicin conjugate-hybridized gold nanorod for combined photothermal-chemotherapy," Biomaterials 35(24), 6576–6584 (2014).
- Kateb B, Chiu K, Black KL. (2011). Nanoplatforms for constructing new approaches to cancer treatment, imaging, and drug delivery:What should be the policy?. Neuro Image, 54(1), S106–S124.
- X. Yang, L. Zhang, and X. Jiang, "Aminosaccharidegold nanoparticle assemblies as narrow-spectrum antibiotics against methicillin-resistant Staphylococcus aureus," Nano Res. 11(12), 6237–6243 (2018).
- A. Ahmed, A. K. Khan, A. Anwar, S. A. Ali, and M. R. Shah, "Biofilm inhibitory effect of chlorhexidine conjugated gold nanoparticles against Klebsiella pneumoniae," Microb. Pathog. 98, 50–56 (2016).
- M. Shilo, P. Berenstein, T. Dreifuss, Y. Nash, G. Goldsmith, G. Kazimirsky, M. Motiei, D. Frenkel, C. Brodie, and R. Popovtzer, "Insulin-coated gold

nanoparticles as a new concept for personalized and adjustable glucose regulation," Nanoscale 7(48), 20489–20496 (2015).

- Lan M-Y, Hsu Y-B, Hsu C-H, Ho C-Y, Lin J-C, Lee S-W. Induction of apoptosis by high-dose gold nanoparticles in nasopharyngeal carcinoma cells. Auris Nasus Larynx 2013; 40: 563-568.
- N. Rattanata, S. Klaynongsruang, C. Leelayuwat, T. Limpaiboon, A. Lulitanond, P. Boonsiri, S. Chio-Srichan, S. Soontaranon, S. Rugmai, and J. Daduang, "Gallic acid conjugated with gold nanoparticles: antibacterial activity and mechanism of action on foodborne pathogens," Int. J. Nanomedicine 11(11), 3347–3356 (2016).
- S. Hayat, S. Muzammil, B. Shabana, B. Aslam, M. H. Siddique, M. Saqalein, and M. A. Nisar, "Quorum quenching: role of nanoparticles as signal jammers in Gram-negative bacteria," Future Microbiol. 14(1), 61– 72 (2019).
- Lukianova-Hleb EY, Wagner DS, Brenner MK, Lapotko DO. Cell-specific transmembrane injection of molecular cargo with gold nanoparticle-generated transient plasmonic nanobubbles. Biomater 2012; 33: 5441-5450.
- 79. V. K. Sonu, I. Rajkumar, K. Bhattacharjee, S. R. Joshi, and S. Mitra, "Interaction of caffeine and sulfadiazine with lysozyme adsorbed at colloidal metal nanoparticle interface: influence on drug transport ability and antibacterial activity," J. Biomol. Struct. Dyn. 37(2), 321–335 (2019).
- M. Imran, A. Hameed, R. M. Shafiullah, R. M. Hafizur, I. Ali, T. Roome, and M. R. Shah, "Fabrication of Xanthan stabilized green gold nanoparticles based tolbutamide delivery system for enhanced insulin secretion in mice pancreatic islets," J. Macromol. Sci. A 55(11–12), 729–735 (2018).
- D. Zhang, J. Zhang, J. Zeng, Z. Li, H. Zuo, C. Huang, and X. Zhao, "Nano-gold loaded with resveratrol enhance the anti-hepatoma effect of resveratrol in vitro and in vivo," J. Biomed. Nanotechnol. 15(2), 288–300 (2019).
- C. Wang, Y. Wang, L. Zhang, R. J. Miron, J. Liang, M. Shi, W. Mo, S. Zheng, Y. Zhao, and Y. Zhang, "Pretreated macrophage-membrane-coated gold nanocages for precise drug delivery for treatment of bacterial infections," Adv. Mater. 30(46), e1804023 (2018).
- 83. Tingting Wang, Yang Jiao, Qinyuan Chai and Xinjun Yu, Gold Nanoparticles: Synthesis and Biological Applications, World Scientific, Vol. 5, No. 3 (2015).
- R. Mendes, A. R. Fernandes, and P. V. Baptista, "Gold nanoparticle approach to the selective delivery of gene silencing in cancer—the case for combined delivery?" Genes (Basel) 8(3), 94 (2017).
- S. Wang, C. Yan, X. Zhang, D. Shi, L. Chi, G. Luo, and J. Deng, "Antimicrobial peptide modification enhances the gene delivery and bactericidal efficiency of gold nanoparticles for accelerating diabetic wound healing," Biomater. Sci. 6(10), 2757–2772 (2018).

- 86. S. N. Barnaby, A. Lee, and C. A. Mirkin, "Probing the inherent stability of siRNA immobilized on nanoparticle constructs," Proc. Natl. Acad. Sci. U.S.A. 111(27), 9739–9744 (2014).
- A. Artiga, I. Serrano-Sevilla, L. De Matteis, S. G. Mitchell, and J. M. de la Fuente, "Current status and future perspectives of gold nanoparticle vectors for siRNA delivery," J. Mater. Chem. B Mater. Biol. Med. 7(6), 876–896 (2019).
- E. Morgan, D. Wupperfeld, D. Morales, and N. Reich, "Shape matters: Gold nanoparticle shape impacts the biological activity of siRNA delivery," Bioconjug. Chem. 30(3), 853–860 (2019).
- R. Calderon-Gonzalez, H. Terán-Navarro, I. García, M. Marradi, D. Salcines-Cuevas, S. Yañez-Diaz, A. Solis- Angulo, E. Frande-Cabanes, M. C. Fariñas, A. Garcia-Castaño, J. Gomez-Roman, S. Penades, F. Rivera, J. Freire, and C. Álvarez-Domínguez, "Gold glyconanoparticles coupled to listeriolysin O 91-99 peptide serve as adjuvant therapy against melanoma," Nanoscale 9(30), 10721–10732 (2017).
- L. J. Cruz, F. Rueda, B. Cordobilla, L. Simón, L. Hosta, F. Albericio, and J. C. Domingo, "Targeting nanosystems to human DCs via Fc receptor as an effective strategy to deliver antigen for immunotherapy," Mol. Pharm. 8(1), 104–116 (2011).
- 91. E. C. Dreaden, S. C. Mwakwari, L. A. Austin, M. J. Kieffer, A. K. Oyelere, and M. A. El-Sayed, "Small molecule-gold nanorod conjugates selectively target and induce macrophage cytotoxicity towards breast cancer cells," Small 8(18), 2819–2822 (2012).
- K. Lee, M. Conboy, H. M. Park, F. Jiang, H. J. Kim, M. A. Dewitt, V. A. Mackley, K. Chang, A. Rao, C. Skinner, T. Shobha, M. Mehdipour, H. Liu, W. C. Huang, F. Lan, N. L. Bray, S. Li, J. E. Corn, K. Kataoka, J. A. Doudna, I. Conboy, and N. Murthy, "Nanoparticle delivery of Cas9 ribonucleoprotein and donor DNA in vivo induces homology-directed DNA repair," Nat. Biomed. Eng. 1(11), 889–901 (2017).
- R. Mout, M. Ray, G. Yesilbag Tonga, Y. W. Lee, T. Tay, K. Sasaki, and V. M. Rotello, "Direct cytosolic delivery of CRISPR/Cas9-ribonucleoprotein for efficient gene editing," ACS Nano 11(3), 2452–2458 (2017).
- 94. M. Schomaker, D. Killian, S. Willenbrock, D. Heinemann, S. Kalies, A. Ngezahayo, I. Nolte, T. Ripken, C. Junghanß, H. Meyer, H. Murua Escobar, and A. Heisterkamp, "Biophysical effects in off-resonant gold nanoparticle mediated (GNOME) laser transfection of cell lines, primary- and stem cells using fs laser pulses," J. Biophotonics 8(8), 646–658 (2015).
- 95. R. Xiong, K. Raemdonck, K. Peynshaert, I. Lentacker, I. De Cock, J. Demeester, S. C. De Smedt, A. G. Skirtach, and K. Braeckmans, "Comparison of gold nanoparticle mediated photoporation: vapor nanobubbles outperform direct heating for delivering macromolecules in live cells," ACS Nano 8(6), 6288– 6296 (2014).
- 96. H. Nakatsuji, K. Kawabata Galbraith, J. Kurisu, H. Imahori, T. Murakami, and M. Kengaku, "Surface

chemistry for cytosolic gene delivery and photothermal transgene expression by gold nanorods," Sci. Rep. 7(1), 4694 (2017).

- C. Yao, F. Rudnitzki, G. Hüttmann, Z. Zhang, and R. Rahmanzadeh, "Important factors for cell-membrane permeabilization by gold nanoparticles activated by nanosecond-laser irradiation," Int. J. Nanomedicine 12(12), 5659–5672 (2017).
- T. Pylaev, E. Vanzha, E. Avdeeva, B. Khlebtsov, and N. Khlebtsov, "A novel cell transfection platform based on laser optoporation mediated by Au nanostar layers," J. Biophotonics 12(1), (2019).
- Z. Lyu, F. Zhou, Q. Liu, H. Xue, Q. Yu, and H. Chen, "A universal platform for macromolecular deliveryinto cells using gold nanoparticle layers via the photoporation effect," Adv. Funct. Mater. 26(32), 5787–5795 (2016).
- 100. J. Hühn, C. Carrillo-Carrion, M. G. Soliman, C. Pfeiffer, D. Valdeperez, A. Masood, I. Chakraborty, L. Zhu, M. Gallego, Z. Yue, M. Carril, N. Feliu, A. Escudero, A. M. Alkilany, B. Pelaz, P. del Pino, and W. J. Parak, "Selected standard protocols for the synthesis, phase transfer, and characterization of inorganic colloidal nanoparticles," Chem. Mater. 29(1), 399–461 (2017).
- 101. Guo Q, Guo Q, Yuan J, Zeng J. Biosynthesis of gold nanoparticles using a kind of flavonol: Dihydromyricetin. Colloids and Surfaces A: Physicochemical and Engineering Aspects 2014; 441: 127-132.
- 102. K. de Oliveira Gonçalves, M. N. da Silva, L. B. Sicchieri, F. R. de Oliveira Silva, R. A. de Matos, and L. C. Courrol, "Aminolevulinic acid with gold nanoparticles: a novel theranostic agent for atherosclerosis," Analyst (Lond.) 140(6), 1974–1980 (2015).
- 103. M. Khoobchandani, K. Katti, A. Maxwell, W. P. Fay, and K. V. Katti, "Laminin receptor-avid nanotherapeutic EGCg-AuNPs as a potential alternative therapeutic approach to prevent restenosis," Int. J. Mol. Sci. 17(3), 316 (2016).
- 104. G. Li, D. Li, L. Zhang, J. Zhai, and E. Wang, "Onestep synthesis of folic acid protected gold nanoparticles and their receptor-mediated intracellular uptake," Chemistry 15(38), 9868–9873 (2009).
- 105. O. Bibikova, P. Singh, A. Popov, G. Akchurin, A. Skaptsov, I. Skovorodkin, V. Khanadeev, D. Mikhalevich, M. Kinnunen, G. Akchurin, V. Bogatyrev, N. Khlebtsov, S. J. Vainio, I. Meglinski, and V. Tuchin, "Shapedependent interaction of gold nanoparticles with cultured cells at laser exposure," Laser Phys. Lett. 14(5), 055901 (2017).
- 106. S.-M. Lee, H. Park, and K.-H. Yoo, "Synergistic cancer therapeutic effects of locally delivered drug and heat using multifunctional nanoparticles," Adv. Mater. 22(36), 4049–4053 (2010).
- 107. R. Chen, X. Zheng, H. Qian, X. Wang, J. Wang, and X. Jiang, "Combined near-IR photothermal therapy and chemotherapy using gold-nanorod/chitosan hybrid

nanospheres to enhance the antitumor effect," Biomater. Sci. 1(3), 285–293 (2013).

- 108. Giljohann DA, Seferos DS, Daniel WL, Massich MD, Patel PC, Mirkin CA. Gold nanoparticles for biology and medicine. Angewandte Chem 2010; 49: 3280-3294.
- 109. S. Gurunathan and J.-H. Kim, "Biocompatible gold nanoparticles ameliorate retinoic acid-induced cell death and induce differentiation in F9 teratocarcinoma stem cells," Nanomaterials (Basel) 8(6), 396 (2018).
- 110. J. N. Payne, H. K. Waghwani, M. G. Connor, W. Hamilton, S. Tockstein, H. Moolani, F. Chavda, V. Badwaik, M. B. Lawrenz, and R. Dakshinamurthy, "Novel synthesis of kanamycin conjugated gold nanoparticles with potent antibacterial activity," Front. Microbiol. 7, 607 (2016).
- 111. H.-Z. Lai, W.-Y. Chen, C.-Y. Wu, and Y.-C. Chen, "Potent antibacterial nanoparticles for pathogenic bacteria," ACS Appl. Mater. Interfaces 7(3), 2046– 2054 (2015).
- 112. Q. You, X. Zhang, F.-G. Wu, and Y. Chen, "Colorimetric and test stripe-based assay of bacteria by using vancomycin-modified gold nanoparticles," Sens. Actuators B Chem. 281, 408–414 (2019).
- 113. M. Demurtas and C. C. Perry, "Facile one-pot synthesis of amoxicillin-coated gold nanoparticles and their antimicrobial activity," Gold Bull. 47(1–2), 103– 107 (2014).
- 114. M. J. Silvero C, D. M. Rocca, E. A. de la Villarmois, K. Fournier, A. E. Lanterna, M. F. Pérez, M. C. Becerra, and J. C. Scaiano, "Selective photoinduced antibacterial activity of amoxicillin-coated gold nanoparticles: from one-step synthesis to in vivo cytocompatibility," ACS Omega 3(1), 1220–1230 (2018).
- 115. M. B. Haddada, K. Jeannot, and J. Spadavecchia, "Novel synthesis and characterization of doxycyclineloaded gold nanoparticles: the golden doxycycline for antibacterial applications," Part. Part. Syst. Charact. 36(2), 1800395 (2019).
- 116. A. Anwar, R. Siddiqui, M. Raza Shah, and N. Ahmed Khan, "Gold nanoparticles conjugation enhances antiacanthamoebic properties of nystatin, fluconazole and amphotericin B," J. Microbiol. Biotechnol. 29(1), 171–177 (2019).
- 117. H. Qiu, Y. Min, Z. Rodgers, L. Zhang, and A. Z. Wang, "Nanomedicine approaches to improve cancer immunotherapy," Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol. 9(5), e1456 (2017).
- 118. A. M. Grimaldi, M. Incoronato, M. Salvatore, and A. Soricelli, "Nanoparticle-based strategies for cancer immunotherapy and immunodiagnostics," Nanomedicine (Lond.) 12(19), 2349–2365 (2017).
- 119. X. Hu, T. Wu, Y. Bao, and Z. Zhang, "Nanotechnology based therapeutic modality to boost anti-tumor immunity and collapse tumor defense," J. Control. Release 256, 26–45 (2017).
- 120. R. Mahjub, S. Jatana, S. E. Lee, Z. Qin, G. Pauli, M. Soleimani, S. Madadi, and S.-D. Li, "Recent advances

in applying nanotechnologies for cancer immunotherapy," J. Control. Release 288, 239–263 (2018).

- 121. Etame AB, Smith CA, Chan WC, Rutka JT. Design and potential application of PEGylated gold nanoparticles with size-dependent permeation through brain microvasculature. Nanomed: Nanotechnol Biol Med 2011; 7: 992-1000.
- 122. H. Y. Yoon, S. T. Selvan, Y. Yang, M. J. Kim, D. K. Yi, I. C. Kwon, and K. Kim, "Engineering nanoparticle strategies for effective cancer immunotherapy," Biomaterials 178, 597–607 (2018).
- 123. J. Jia, Y. Zhang, Y. Xin, C. Jiang, B. Yan, and S. Zhai, "Interactions between nanoparticles and dendritic cells: from the perspective of cancer immunotherapy," Front. Oncol. 8, 404 (2018).
- 124. B. Englinger, C. Pirker, P. Heffeter, A. Terenzi, C. R. Kowol, B. K. Keppler, and W. Berger, "Metal drugs and the anticancer immune response," Chem. Rev. 119(2), 1519–1624 (2019).
- 125. Jeff MMJM, Bulte WM. (2008). Nanoparticles in biomedical imaging, emerging technologies and applications. Fundamental Biomedical Technologies, Springer, New York, NY, USA, 102.
- 126. F. Lopez-Campos, D. Candini, E. Carrasco, and M. A. Berenguer Francés, "Nanoparticles applied to cancer immunoregulation," Rep. Pract. Oncol. Radiother. 24(1), 47–55 (2019).
- 127. Q.-V. Le, G. Yang, Y. Wu, H. W. Jang, M. Shokouhimehr, and Y.-K. Oh, "Nanomaterials for modulating innate immune cells in cancer immunotherapy," Asian J. Pharm. Sci. 14(1), 16–29 (2019).
- 128. L. A. Dykman, S. A. Staroverov, A. S. Fomin, V. A. Khanadeev, B. N. Khlebtsov, and V. A. Bogatyrev, "Gold nanoparticles as an adjuvant: Influence of size, shape, and technique of combination with CpG on antibody production," Int. Immunopharmacol. 54, 163–168 (2018).
- 129. R. Meir, K. Shamalov, T. Sadan, M. Motiei, G. Yaari, C. J. Cohen, and R. Popovtzer, "Fast image-guided stratification using anti-programmed death ligand 1 gold nanoparticles for cancer immunotherapy," ACS Nano 11(11), 11127–11134 (2017).
- 130. F. Sousa, P. Castro, P. Fonte, P. J. Kennedy, M. T. Neves-Petersen, and B. Sarmento, "Nanoparticles for the delivery of therapeutic antibodies: Dogma or promising strategy?" Expert Opin. Drug Deliv. 14(10), 1163–1176 (2017).
- 131. Y.-S. S. Yang, K. D. Moynihan, A. Bekdemir, T. M. Dichwalkar, M. M. Noh, N. Watson, M. Melo, J. Ingram, H. Suh, H. Ploegh, F. R. Stellacci, and D. J. Irvine, "Targeting small molecule drugs to T cells with antibodydirected cell-penetrating gold nanoparticles," Biomater. Sci. 7(1), 113–124 (2018).
- 132. I. Mottas, A. Bekdemir, A. Cereghetti, L. Spagnuolo, Y.-S. S. Yang, M. Müller, D. J. Irvine, F. Stellacci, and C. Bourquin, "Amphiphilic nanoparticle delivery enhances the anticancer efficacy of a TLR7 ligand via

local immune activation," Biomaterials 190-191, 111–120 (2019).

- 133. J. P. M. Almeida, A. Y. Lin, R. J. Langsner, P. Eckels, A. E. Foster, and R. A. Drezek, "In vivo immune cell distribution of gold nanoparticles in naïve and tumor bearing mice," Small 10(4), 812–819 (2014).
- 134. S. Saha, X. Xiong, P. K. Chakraborty, K. Shameer, R. R. Arvizo, R. A. Kudgus, S. K. D. Dwivedi, M. N. Hossen, E. M. Gillies, J. D. Robertson, J. T. Dudley, R. A. Urrutia, R. G. Postier, R. Bhattacharya, and P. Mukherjee, "Gold nanoparticle reprograms pancreatic tumor microenvironment and inhibits tumor growth," ACS Nano 10(12), 10636–10651 (2016).
- 135. Y. S. Tsai, Y. H. Chen, P. C. Cheng, H. T. Tsai, A. L. Shiau, T. S. Tzai, and C. L. Wu, "TGF-β1 conjugated to gold nanoparticles results in protein conformational changes and attenuates the biological function," Small 9(12), 2119–2128 (2013).
- 136. A. Y. Lin, J. Lunsford, A. S. Bear, J. K. Young, P. Eckels, L. Luo, A. E. Foster, and R. A. Drezek, "High-density sub-100-nm peptide-gold nanoparticle complexes improve vaccine presentation by dendritic cells in vitro," Nanoscale Res. Lett. 8(1), 72 (2013).
- 137. J. Yu, X. Wang, Y. Li, X. Huang, X. Luo, and X. He, "Synthesis of nerolidol functionalized gold nanoparticles for wound regeneration in people with diabetic foot ulcers in nursing care management," Sci. Adv. Mater. 10(12), 1775–1781 (2018).
- 138. S. K. Vemuri, R. R. Banala, S. Mukherjee, P. Uppula, G. Subbaiah, R.G.V. Gurava, and T. Malarvilli, "Novel biosynthesized gold nanoparticles as anti-cancer agents against breast cancer: Synthesis, biological evaluation, molecular modelling studies," Mater. Sci. Eng. C 99, 417–429 (2019).
- 139. H. Deng, F. Dai, G. Ma, and X. Zhang, "Theranostic gold nanomicelles made from biocompatible comblike polymers for thermochemotherapy and multifunctional imaging with rapid clearance," Adv. Mater. 27(24), 3645–3653 (2015).
- 140. Y. M. Park, S. J. Lee, Y. S. Kim, M. H. Lee, G. S. Cha, I. D. Jung, T. H. Kang, and H. D. Han, "Nanoparticlebased vaccine delivery for cancer immunotherapy," Immune Netw. 13(5), 177–183 (2013).
- 141. Heo DN, Yang DH, Moon HJ. (2012). Gold nanoparticles surface-functionalized with paclitaxel drug and biotin receptor as theranostic agents for cancer therapy. Biomaterials, 33(3), 856- 866.
- 142. S. Biswas, S. H. Medina, and J. J. Barchi, Jr., "Synthesis and cell-selective antitumor properties of amino acid conjugated tumor-associated carbohydrate antigencoated gold nanoparticles," Carbohydr. Res. 405, 93– 101 (2015).
- 143. Q. Zhou, Y. Zhang, J. Du, Y. Li, Y. Zhou, Q. Fu, J. Zhang, X. Wang, and L. Zhan, "Different-sized gold nanoparticle activator/antigen increases dendritic cells accumulation in liver-draining lymph nodes and Cd8+ T cell responses," ACS Nano 10(2), 2678–2692 (2016).

- 144. X. Ma, H. Hui, Y. Jin, D. Dong, X. Liang, X. Yang, K. Tan, Z. Dai, Z. Cheng, and J. Tian, "Enhanced immunotherapy of SM5-1 in hepatocellular carcinoma by conjugating with gold nanoparticles and its in vivo bioluminescence tomographic evaluation," Biomaterials 87, 46–56 (2016).
- 145. L. A. Dykman, M. V. Sumaroka, S. A. Staroverov, I. S. Zaĭtseva, and V. A. Bogatyrev, "[Immunogenic properties of the colloidal gold]," Izv. Akad. Nauk Ser. Biol. 31(1), 86–91 (2004).
- 146. S. Ahn, I. H. Lee, S. Kang, D. Kim, M. Choi, P. E. Saw, E. C. Shin, and S. Jon, "Gold nanoparticles displaying tumor-associated self-antigens as a potential vaccine for cancer immunotherapy," Adv. Healthc. Mater. 3(8), 1194–1199 (2014).
- 147. S. Tomić, J. Đokić, S. Vasilijić, N. Ogrinc, R. Rudolf, P. Pelicon, D. Vučević, P. Milosavljević, S. Janković, I. Anžel, J. Rajković, M. S. Rupnik, B. Friedrich, and M. Colić, "Size-dependent effects of gold nanoparticles uptake on maturation and antitumor functions of human dendritic cells in vitro," PLoS One 9(5), e96584 (2014).
- 148. S. Kang, S. Ahn, J. Lee, J. Y. Kim, M. Choi, V. Gujrati, H. Kim, J. Kim, E.-C. Shin, and S. Jon, "Effects of gold nanoparticle-based vaccine size on lymph node delivery and cytotoxic T-lymphocyte responses," J. Control. Release 256, 56–67 (2017).
- 149. C. Wu, H. Chen, X. Wu, X. Cong, L. Wang, Y. Wang, Y. Yang, W. Li, and T. Sun, "The influence of tumorinduced immune dysfunction on the immune cell distribution of gold nanoparticles in vivo," Biomater. Sci. 5(8), 1531–1536 (2017).
- 150. H. A. Andersson, Y.-S. Kim, B. E. O'Neill, Z.-Z. Shi, and R. E. Serda, "HSP70 promoter-driven activation of gene expression for immunotherapy using gold nanorods and near infrared light," Vaccines (Basel) 2(2), 216–227 (2014).
- 151. E. de Alteriis, V. Maselli, A. Falanga, S. Galdiero, F. M. Di Lella, R. Gesuele, M. Guida, and E. Galdiero, "Efficiency of gold nanoparticles coated with the antimicrobial peptide indolicidin against biofilm formation and development of Candida spp. clinical isolates," Infect. Drug Resist. 11(11), 915–925 (2018).
- 152. R. Liang, J. Xie, J. Li, K. Wang, L. Liu, Y. Gao, M. Hussain, G. Shen, J. Zhu, and J. Tao, "Liposomescoated gold nanocages with antigens and adjuvants targeted delivery to dendritic cells for enhancing antitumor immune response," Biomaterials 149, 41– 50 (2017).
- 153. Rastogi L, Kora AJ, J A. Highly stable, protein capped gold nanoparticles as effective drug delivery vehicles for amino-glycosidic antibiotics. Mater Sci Engineer: C 2012; 32: 1571-1577.
- 154. S. Ahmad, A. A. Zamry, H.-T. T. Tan, K. K. Wong, J. Lim, and R. Mohamud, "Targeting dendritic cells through gold nanoparticles: A review on the cellular uptake and subsequent immunological properties," Mol. Immunol. 91, 123–133 (2017).
- 155. S. Fogli, C. Montis, S. Paccosi, A. Silvano, E. Michelucci, D. Berti, A. Bosi, A. Parenti, and P.

Romagnoli, "Inorganic nanoparticles as potential regulators of immune response in dendritic cells," Nanomedicine (Lond.) 12(14), 1647–1660 (2017).

- 156. Hartono D, Hody, Yang KL, Yung LY. The effect of cholesterol on protein-coated gold nanoparticle binding to liquid crystal-supported models of cell membranes. Biomater 2010; 31: 3008-3015.
- 157. Gao W, Xu K, Ji L, Tang B. Effect of gold nanoparticles on glutathione depletion-induced hydrogen peroxide generation and apoptosis in HL7702 cells. Toxicol Lett. 2011;205(1):86–95.
- 158. Arvizo R, Bhattacharya R, Mukherjee P. Gold nanoparticles : Opportunities and Challenges in Nanomedicine Gold nanoparticles : opportunities and challenges in nanomedicine. 2010; (May 2014).
- 159. Agasti SS, Chompoosor A, You CC, Ghosh P, Kim CK, Rotello VM. (2009). Photoregulated release of caged anticancer drugs from gold nanoparticles. Journal of the American Chemical Society, 131(16), 5728–5729.
- 160. X. Wang, J. Li, N. Kawazoe, and G. Chen, "Photothermal ablation of cancer cells by albuminmodified gold nanorods and activation of dendritic cells," Materials (Basel) 12(1), 31 (2018).
- 161. M. R. Choi, K. J. Stanton-Maxey, J. K. Stanley, C. S. Levin, R. Bardhan, D. Akin, S. Badve, J. Sturgis, J. P. Robinson, R. Bashir, N. J. Halas, and S. E. Clare, "A cellular Trojan Horse for delivery of therapeutic nanoparticles into tumors," Nano Lett. 7(12), 3759– 3765 (2007).
- 162. E. Spyratou, M. Makropoulou, E. P. Efstathopoulos, A. G. Georgakilas, and L. Sihver, "Recent advances in cancer therapy based on dual mode gold nanoparticles," Cancers (Basel) 9(12), 173 (2017).
- 163. Mendoza KC, McLane VD, Kim S, Griffin JD. Invitro application of gold nanoprobes in live neurons for phenotypical classification, connectivity assessment, and electrophysiological recording. Brain Res 2010; 1325: 19-27.
- 164. Y. Li, X. Li, F. Zhou, A. Doughty, A. R. Hoover, R. E. Nordquist, and W. R. Chen, "Nanotechnology-based photoimmunological therapies for cancer," Cancer Lett. 442, 429–438 (2019).
- 165. J. Beik, M. Khateri, Z. Khosravi, S. K. Kamrava, S. Kooranifar, H. Ghaznavi, and A. Shakeri-Zadeh, "Gold nanoparticles in combinatorial cancer therapy strategies," Coord. Chem. Rev. 387, 299–324 (2019).
- 166. J. You, R. Zhang, G. Zhang, M. Zhong, Y. Liu, C. S. Van Pelt, D. Liang, W. Wei, A. K. Sood, and C. Li, "Photothermal-chemotherapy with doxorubicinloaded hollow gold nanospheres: A platform for nearinfrared light-trigged drug release," J. Control. Release 158(2), 319–328 (2012).
- 167. R. Mendes, P. Pedrosa, J. C. Lima, A. R. Fernandes, and P. V. Baptista, "Photothermal enhancement of chemotherapy in breast cancer by visible irradiation of Gold Nanoparticles," Sci. Rep. 7(1), 10872 (2017).
- 168. J. Nam, S. Son, L. J. Ochyl, R. Kuai, A. Schwendeman, and J. J. Moon, "Chemo-photothermal therapy combination elicits anti-tumor immunity against

advanced metastatic cancer," Nat. Commun. 9(1), 1074 (2018).

- 169. G. Bisker, D. Yeheskely-Hayon, L. Minai, and D. Yelin, "Controlled release of Rituximab from gold nanoparticles for phototherapy of malignant cells," J. Control. Release 162(2), 303–309 (2012).
- 170. J. Shao, R. J. Griffin, E. I. Galanzha, J. W. Kim, N. Koonce, J. Webber, T. Mustafa, A. S. Biris, D. A. Nedosekin, and V. P. Zharov, "Photothermal nanodrugs: potential of TNF-gold nanospheres for cancer theranostics," Sci. Rep. 3(1), 1293 (2013).
- 171. J. Nam, W.-G. La, S. Hwang, Y. S. Ha, N. Park, N. Won, S. Jung, S. H. Bhang, Y.-J. Ma, Y.-M. Cho, M. Jin, J. Han, J.-Y. Shin, E. K. Wang, S. G. Kim, S.-H. Cho, J. Yoo, B.-S. Kim, and S. Kim, "pH-responsive assembly of gold nanoparticles and "spatiotemporally concerted" drug release for synergistic cancer therapy," ACS Nano 7(4), 3388–3402 (2013).
- 172. Y. Tao, E. Ju, Z. Liu, K. Dong, J. Ren, and X. Qu, "Engineered, self-assembled near-infrared photothermal agents for combined tumor immunotherapy and chemo-photothermal therapy," Biomaterials 35(24), 6646–6656 (2014).
- 173. B. K. Poudel, B. Gupta, T. Ramasamy, R. K. Thapa, S. Pathak, K. T. Oh, J. H. Jeong, H. G. Choi, C. S. Yong, and J. O. Kim, "PEGylated thermosensitive lipid-coated hollow gold nanoshells for effective combinational chemo-photothermal therapy of pancreatic cancer," Colloids Surf. B Biointerfaces 160, 73–83 (2017).
- 174. H. Kim, V. P. Nguyen, P. Manivasagan, M. J. Jung, S. W. Kim, J. Oh, and H. W. Kang, "Doxorubicinfucoidangold nanoparticles composite for dualchemo-photothermal treatment on eye tumors," Oncotarget 8(69), 113719 (2017).
- 175. L. Liu, H. J. Xie, L. M. Mu, R. Liu, Z. B. Su, Y. N. Cui, Y. Xie, and W. L. Lu, "Functional chlorin gold nanorods enable to treat breast cancer by photothermal/photodynamic therapy," Int. J. Nanomedicine 13(13), 8119–8135 (2018).
- 176. T. H. Tran, R. K. Thapa, H. T. Nguyen, T. T. Pham, T. Ramasamy, D. S. Kim, C. S. Yong, J. O. Kim, and H.-G. Choi, "Combined phototherapy in anti-cancer treatment: therapeutics design and perspectives," J. Pharm. Investig. 46(6), 505–517 (2016).
- 177. F. Cao, M. Yan, Y. Liu, L. Liu, and G. Ma, "Photothermally controlled MHC class I restricted CD8+ T-cell responses elicited by hyaluronic acid decorated gold nanoparticles as a vaccine for cancer immunotherapy," Adv. Healthc. Mater. 7(10), e1701439 (2018).
- 178. B. Zhou, J. Song, M. Wang, X. Wang, J. Wang, E. W. Howard, F. Zhou, J. Qu, and W. R. Chen, "BSAbioinspired gold nanorods loaded with immunoadjuvant for the treatment of melanoma by combined photothermal therapy and immunotherapy," Nanoscale 10(46), 21640–21647 (2018).
- 179. J. Chen, L. Lin, N. Yan, Y. Hu, H. Fang, Z. Guo, P. Sun, H. Tian, and X. Chen, "Macrophages loaded CpG

and GNR-PEI for combination of tumor photothermal therapy and immunotherapy," Sci. China Mater. 61(11), 1484–1494 (2018).

- 180. D. Chu, X. Dong, Q. Zhao, J. Gu, and Z. Wang, "Photosensitization priming of tumor microenvironments improves delivery of nanotherapeutics via neutrophil infiltration," Adv. Mater. 29(27), 1701021 (2017).
- 181. L. Au, D. Zheng, F. Zhou, Z. Y. Li, X. Li, and Y. Xia, "A quantitative study on the photothermal effect of immuno gold nanocages targeted to breast cancer cells," ACS Nano 2(8), 1645–1652 (2008).
- 182. M. Zhang, H. S. Kim, T. Jin, J. Woo, Y. J. Piao, and W. K. Moon, "Near-infrared photothermal therapy using anti-EGFR-gold nanorod conjugates for triple negative breast cancer," Oncotarget 8(49), 86566–86575 (2017).
- 183. P. Vijayaraghavan, C. H. Liu, R. Vankayala, C. S. Chiang, and K. C. Hwang, "Designing multi-branched gold nanoechinus for NIR light activated dual modal photodynamic and photothermal therapy in the second biological window," Adv. Mater. 26(39), 6689– 6695 (2014).
- 184. A. Barhoumi, W. Wang, D. Zurakowski, R. S. Langer, and D. S. Kohane, "Photothermally targeted thermosensitive polymer-masked nanoparticles," Nano Lett. 14(7), 3697–3701 (2014).
- 185. Z. Wang, S. Li, M. Zhang, Y. Ma, Y. Liu, W. Gao, J. Zhang, and Y. Gu, "Laser-triggered small interfering RNA releasing gold nanoshells against heat shock protein for sensitized photothermal therapy," Adv. Sci. (Weinh.) 4(2), 1600327 (2016).
- 186. D. G. Meeker, T. Wang, W. N. Harrington, V. P. Zharov, S. A. Johnson, S. V. Jenkins, S. E. Oyibo, C. M. Walker, W. B. Mills, M. E. Shirtliff, K. E. Beenken, J. Chen, and M. S. Smeltzer, "Versatility of targeted antibiotic-loaded gold nanoconstructs for the treatment of biofilm-associated bacterial infections," Int. J. Hyperthermia 34(2), 209–219 (2018).
- 187. Q. Chen, L. Zhang, Y. Feng, F. Shi, Y. Wang, P. Wang, and L. Liu, "Dual-functional peptide conjugated gold nanorods for the detection and photothermal ablation of pathogenic bacteria," J. Mater. Chem. B Mater. Biol. Med. 6(46), 7643–7651 (2018).
- 188. Y. Zhao, Q. Guo, X. Dai, X. Wei, Y. Yu, X. Chen, C. Li, Z. Cao, and X. Zhang, "A biomimetic non-antibiotic approach to eradicate drug-resistant infections," Adv. Mater. 31(7), e1806024 (2019).
- 189. M. Zhang, T. Yilmaz, A. O. Boztas, O. Karakuzu, W. Y. Bang, Y. Yegin, Z. Luo, M. Lenox, L. Cisneros-Zevallos, and M. Akbulut, "A multifunctional nanoparticulate theranostic system with simultaneous chemotherapeutic, photothermal therapeutic, and MRI contrast capabilities," RSC Advances 6(33), 27798–27806 (2016).
- 190. Chen WH, Xu XD, Jia HZ. (2013). Therapeutic nanomedicine based on dual-intelligent functionalized gold nanoparticles for cancer imaging and therapyin vivo. Biomaterials. 34(34), 8798- 8807.

- 191. N. Amreddy, R. Muralidharan, A. Babu et al., "Tumor-targeted and pH-controlled delivery of doxorubicin using gold nanorods for lung cancer therapy," International Journal of Nanomedicine, vol. 10, pp. 6773–6788, 2015.
- 192. A. Hatef, S. Fortin-Desch<sup>enes</sup>, E. Boulais, F. Lesage, and M. Meunier, "Photothermal response of hollow gold nanoshell to laser irradiation: continuous wave, short and ultrashort pulse," International Journal ofHeat andMass Transfer, vol. 89, pp. 866–871, 2015.
- 193. C. Du, A. Wang, J. Fei, J. Zhao, and J. Li, "Polypyrrolestabilized gold nanorodswith enhanced photothermal effect towards twophoton photothermal therapy," Journal of Materials Chemistry B, vol. 3, no. 22, pp. 4539–4545, 2015.
- 194. A. K. Singh, X. Bai, M. A. R. Amalaradjou, and A. K. Bhunia, "Antilisterial and antibiofilm activities of Pediocin and LAP functionalized gold nanoparticles," Front. Sustain. Food Syst. 2, 74,2015.
- 195. C.-C. Huang and T.-M. Liu, "Controlled Au-polymer nanostructures for multiphoton imaging, prodrug delivery, and chemo-photothermal therapy platforms," ACS Appl. Mater. Interfaces 7(45), 25259– 25269 (2015).
- 196. Z. Zhang, L. Wang, J. Wang, X. Jiang, X. Li, Z. Hu, Y. Ji, X. Wu, and C. Chen, "Mesoporous silica-coated gold nanorods as a light-mediated multifunctional theranostic platform for cancer treatment," Adv. Mater. 24(11), 1418–1423 (2012).
- 197. N. Li, J. Cheng, Y. Zhang, J. Wang, G. Huang, J. Zhu, and D. He, "A chemophotothermal and targeting multifunctional nanoprobe with a tumor-diagnosing ability," Nano Res. 11(8), 4333–4347 (2018).
- 198. W. C. Huang, P.-J. Tsai, and Y.-C. Chen, "Multifunctional Fe3O4@Au nanoeggs as photothermal agents forselective killing of nosocomial and antibiotic-resistant bacteria," Small 5(1), 51–56 (2009).
- 199. M. I. Setyawati, C. Y. Tay, B. H. Bay, and D. T. Leong, "Gold nanoparticles induced endothelial leakiness depends on particle size and endothelial cell origin," ACS Nano 11(5), 5020–5030 (2017).
- 200. C. Y. Tay, M. I. Setyawati, and D. T. Leong, "Nanoparticle density: a critical biophysical regulator of endothelial permeability," ACS Nano 11(3), 2764– 2772 (2017).
- 201. F. Peng, M. I. Setyawati, J. K. Tee, X. Ding, J. Wang, M. E. Nga, H. K. Ho, and D. T. Leong, "Nanoparticles promote in vivo breast cancer cell intravasation and extravasation by inducing endothelial leakiness," Nat. Nanotechnol. 14(3), 279–286 (2019).
- 202. B. N. Khlebtsov, D. N. Bratashov, N. A. Byzova, B. B. Dzantiev, and N. G. Khlebtsov, "SERS-based lateral flow immunoassay of troponin I using gap-enhanced Raman tags," Nano Res. 12(2), 413–420 (2019).
- 203. K.-Q. Lin, J. Yi, J.-H. Zhong, S. Hu, B.-J. Liu, J.-Y. Liu, C. Zong, Z.-C. Lei, X. Wang, J. Aizpurua, R. Esteban, and B. Ren, "Plasmonic photoluminescence for recovering native chemical information from surface-

enhanced Raman scattering," Nat. Commun. 8(1), 14891 (2017).

- 204. X. Huang, X. Peng, Y. Wang, Y. Wang, D. M. Shin, M. A. El-Sayed, and S. Nie, "A reexamination of active and passive tumor targeting by using rod-shaped gold nanocrystals and covalently conjugated peptide ligands," ACS Nano 4(10), 5887–5896 (2010).
- 205. A. Camposeo, L. Persano, R. Manco et al., "Metal-Enhanced Near-Infrared Fluorescence by Micropatterned Gold Nanocages," ACS Nano, vol. 9, no. 10, pp. 10047–10054, 2015.
- 206. Chen H, Zhang X, Dai S. (2013). Multifunctional gold nano star conjugates for tumor imaging and combined photothermal and chemo-therapy. Theranostics, 3(9),633-49.2018).
- 207. S. C.-H. Tsao, J. Wang, Y. Wang, A. Behren, J. Cebon, and M. Trau, "Characterising the phenotypic evolution of circulating tumour cells during treatment," Nat. Commun. 9(1), 1482 (2018).
- 208. Z. Zhang, S. Wang, H. Xu, B. Wang, and C. Yao, "Role of 5-aminolevulinic acid-conjugated gold nanoparticles for photodynamic therapy of cancer," Journal of Biomedical Optics, vol. 20, no. 5, Article ID 051043, 2015.
- 209. K. B. Vang, I. Safina, E. Darrigues, D. Nedosekin, Z. A. Nima, W. Majeed, F. Watanabe, G. Kannarpady, R. A. Kore, D. Casciano, V. P. Zharov, R. J. Griffin, R. P. M. Dings, and A. S. Biris, "Modifying dendritic cell activation with plasmonic nano vectors," Sci. Rep. 7(1), 5513 (2017).
- 210. S. M. R. Safaee, M. Janipour, and M. A. Karami, "Modeling and analysis of optical properties of a gold nanoring based on electric and magnetic dipoles," Applied Optics, vol. 54, no. 28, pp. 8313–8317, 2015.
- Amjadi M, Farzampour L. Fluorescence quenching of fluoroquinolones by gold nanoparticles with different sizes and its analytical application. J Luminesc 2014; 145: 263-268.
- 212. Chakraborty S. (2009). Lipid an emerging platform for oral delivery of drugs with poor bioavailability. European Journal of Pharmaceutics and Biopharmaceutics,73(1), 1-15.
- 213. M. P. Antosh, D.D. Wijesinghe, S. Shrestha et al., "Enhancement of radiation effect on cancer cells by gold-pHLIP," Proceedings of the National Academy of Sciences of the United States of America, vol. 112, no. 17, pp. 5372–5376, 2015.
- Sharma, V., Park, K. & Srinivasarao, M. Colloidal dispersion of Au nanorods: Historical background, optical properties, seed-mediated synthesis, shape separation and self-assembly. Mater. Sci. Eng. R 65, 1-38 (2009).
- 215. Morgan MT, Nakanishi Y, Kroll DJ, Griset AP, Carnahan MA, Wathier M, Oberlies NH, Manikumar G, Wani MC, Grinstaff MW. (2006). Dendrimerencapsulated camptothecins: increased solubility, cellular uptake, and cellular retention affords enhanced anticancer activity in vitro. Cancer Research, 66(24), 11913–11921.

- 216. Liu, X. et al. A one-step homogeneous immunoassay for cancer biomarker detection using Au nanoparticle probes coupled with dynamic light scattering. J. Am. Chem. Soc. 130, 2780-2782 (2008).
- 217. López-García, M.; Galisteo-López, J.F.; Blanco, Á.; López, C.; García-Martín, A. High Degree of Optical Tunability of Self-Assembled Photonic-Plasmonic Crystals by Filling Fraction Modification. Adv. Funct. Mater. 2011, 20, 4338–4343.
- 218. Hong, Y.; Pourmand, M.; Boriskina, S.V.; Reinhard, B.M. Enhanced Light Focusing in Self-Assembled Optoplasmonic Clusters with Subwavelength Dimensions. Adv. Mater. 2012, 25, 115–119.
- 219. Di Guglielmo C, Lopez DR, De Lapuente J, Mallafre JM, Suarez MB. Embryotoxicity of cobalt ferrite and gold nanoparticles: a first in vitro approach. Reproduct Toxicol 2010; 30: 271-276.
- 220. Duncan B, Kim C, Rotello VM. (2010). Gold nanoparticle platforms as drug and biomacromolecule delivery systems. Journal of Controlled Release, 148, 122–127.
- 221. Aaron, J. et al. Polarization microscopy with stellated Au nanoparticles for robust monitoring of molecular assemblies and single biomolecules. Opt. Express. 16, 2153-2167 (2008).
- Zharov, V., Galanzha, E., Shashkov, E., Khlebtsov, N. & Tuchin, V. In vivo photo acoustic flow cytometry for monitoring circulating single cancer cells and contrast agents. Opt. Lett. 31, 3623-3625 (2006).
- 223. Pissuwan, D., Niidome, T. & Cortie, M. B. The forthcoming application of Au nanoparticles in drug and gene delivery systems. J. Control. Release 149, 65-71 (2011).
- 224. Rossi, S.; Donadio, S.; Fontana, L.; Porcaro, F.; Battocchio, C.; Venditti, I.; Bracci, L.; Fratoddi, I. Negatively charged gold nanoparticles as dexamethasone carrier: Stability and citotoxic activity. RCS Adv. 2016, 6, 99016–99022.
- 225. Zhou, J., Ralston, J., Sedev, R. & Beattie, D. A. Functionalized Au nanoparticles: synthesis, structure and colloid stability. J. Colloid Interface Sci. 331, 251-262 (2008).
- 226. Pal R, Panigrahi S, Bhattacharyya D, Chakraborti AS.Characterization of citrate capped gold nanoparticlequercetincomplex: Experimental and quantum chemical approach. J Mol Struct 2013; 1046: 153-163
- 227. Parkhomenko, R.G.; Plekhanov, A.I.; Kuchyanov, A.S.; Trubin, S.V.; Kuchumov, B.M.; Igumenov, I.K. Gold nanostructure formation in the photonic crystal matrix by means of MOCVD technique. Surf. Coat. Technol. 2013, 230, 279–283.
- 228. Venditti, I.; Chronopoulou, L.; Fratoddi, I.; Palocci, C.; Diociaiuti, M.; Russo, M.V. Candida rugosa lipase immobilization on hydrophilic charged gold nanoparticles as promising biocatalysts: Activity and stability investigations. Colloid Surf. B Biointerfaces 2015, 131, 93–101.
- 229. T. Sugiura, D.Matsuki, J. Okajima et al., "Photothermal therapy of tumors in lymph nodes

using gold nanorods and nearinfrared laser light with controlled surface cooling," Nano Research, vol. 8, no. 12, pp. 3842–3852, 2015.

- 230. Cai, Z.; Jun Liu, Y.; Lu, X.; Teng, J. In Situ "Doping" Inverse Silica Opals with Size-Controllable Gold Nanoparticles for Refractive Index Sensing. J. Phys. Chem. C 2013, 117, 9440–9445.
- 231. Kong FY, Zhang JW, Li RF, Wang ZX, Wang WJ, Wang W. Unique roles of gold nanoparticles in drug delivery, targeting and imaging applications.Molecules. 2017;22(9).
- 232. Lan M-Y, Hsu Y-B, Hsu C-H, Ho C-Y, Lin J-C, Lee S-W. Induction of apoptosis by high-dose gold nanoparticles in nasopharyngeal carcinoma cells. Auris Nasus Larynx. 2013;40(6):563–8.
- 233. Hartono D, Yang K-L, Yung L-YL. The effect of cholesterol on protein-coated gold nanoparticle binding to liquid crystal-supported models of cell membranes. Biomaterials. 2010;31(11):3008–15.
- 234. Mishra A, Tripathy SK, Yun S-I. Fungus mediated synthesis of gold nanoparticles and their conjugation with genomic DNA isolated from Escherichia coli and Staphylococcus aureus. Process Biochem. 2012;47(5):701–11.
- 235. Chithrani DB, Dunne M, Stewart J, Allen C, Jaffray DA. Cellular uptake and transport of gold nanoparticles incorporated in a liposomal carrier. Nanomedicine Nanotechnology, Biol Med. 2010;6(1):161–9.
- 236. Di Guglielmo C, López DR, De Lapuente J, Mallafre JML, Suàrez MB. Oxidative stress and toxicity of gold nanoparticles in Mytilus edulis. Reprod Toxicol. 2010;30(2):271–6.
- 237. Wang, D.; Li, J.; Chan, C.T.; Salgueirino-Maceira, V.; LizMarzan, L.M.; Romanov, S.; Caruso, F. Optical Properties of Nanoparticle-Based Metallodielectric Inverse Opals. Small 2005, 1, 122–130
- 238. Di Guglielmo C, López DR, De Lapuente J, Mallafre JML, Suàrez MB. Embryotoxicity of cobalt ferrite and gold nanoparticles: a first in vitro approach. Reprod Toxicol. 2010;30(2):271–6.
- 239. Kojima C, Umeda Y, Harada A, Kono K. Preparation of near-infrared light absorbing gold nanoparticles using polyethylene glycol-attached dendrimers. Colloids surfaces B Biointerfaces. 2010;81(2):648–51.
- 240. Lee K, Lee H, Bae KH, Park TG. Heparin immobilized gold nanoparticles for targeted detection and apoptotic death of metastatic cancer cells. Biomaterials. 2010;31(25):6530–6.
- 241. Tan, Y.; Qian,W.; Ding, S.;Wang, Y. Gold-Nanoparticle Infiltrated Polystyrene InverseOpals: AThree-Dimensional Platform for Generating Combined Optical Properties. Chem. Mater. 2006, 18, 3385–3389.
- 242. J. Li, C. C. Sharkey, D. Huang, and M. R. King, "Nanobiotechnology for the therapeutic targeting of cancer cells in blood," Cellular and Molecular Bioengineering, vol. 8, no. 1, pp. 137–150, 2015.
- 243. Fu, J.; Tandaechanurat, A.; Iwamoto, S.; Arakawa, Y. Design of large-bandwidth single-mode operation

waveguides in silicon three-dimensional photonic crystals using two guided modes. Opt. Express 2013, 21, 12443–12450.

- 244. Wahle, M.; Ebel, J.; Wilkes, D.; Kitzerow, H.-S. Asymmetric band gap shift in electrically addressed blue phase photonic crystal fibers. Opt. Express 2016, 24, 22718–22729.
- 245. Schutzmann, S.; Venditti, I.; Prosposito, P.; Casalboni, M.; Russo, M.V. High-energy angle resolved reflection spectroscopy on three-dimensional photonic crystals of self-organized polymeric nanospheres. Opt. Express 2008, 16, 897–907.
- 246. Miller, O.D.; Polimeridis, A.G.; Homer Reid, M.T.; Hsu, C.W.; DeLacy, B.G.; Joannopoulos, J.D.; Soljacic, M.; Johnson, S.G. Fundamental limits to optical response in absorptive systems. Opt. Express 2016, 24, 3329–3364.
- 247. Lidorikis, E.; Egusa, S.; Joannopoulos, J.D. Effective medium properties and photonic crystal superstructures of metallic nanoparticle arrays. J. Appl. Phys. 2007, 101, 054304.
- 248. Ding, S.; Qian, W.; Tan, Y.; Wang, Y. In-Situ Incorporation of Gold Nanoparticles of Desired Sizes into Three-Dimensional Macroporous Matrixes. Langmuir 2006, 22, 7105–7108.
- 249. Imai, Y.; Finlayson, C.E.; Goldberg-Oppenheimer, P.; Zhao, Q.; Spahn, P.; Snoswell, D.R.E.; Haines, A.I.; Hellmannb, G.P.; Baumberg, J.J. Electrically conductive polymeric photonic crystals. Soft Matter 2012, 8, 6280–6290.
- 250. Galisteo-López, J.F.; Ibisate, M.; Sapienza, R.; Froufe-Pérez, L.S.; Blanco, Á.; López, C. Self-Assembled Photonic Structures. Adv. Mater. 2011, 23, 30–69.
- 251. Bardosova, M.; Pemble, M.E.; Povey, I.M.; Tredgold, R.H. The Langmuir-Blodgett Approach to Making Colloidal Photonic Crystals from Silica Spheres. Adv. Mater. 2010, 22, 3104–3124.
- 252. Jones, M.R.; Osberg, K.D.; Macfarlane, R.J.; Langille, M.R.; Mirkin, C.A. Templated Techniques for the Synthesis and Assembly of Plasmonic Nanostructures. Chem. Rev. 2011, 111, 3736–3827.
- 253. Yu, X.; Shi, L.; Han, D.; Zi, J.; Braun, P.V. High Quality Factor Metallodielectric Hybrid Plasmonic-Photonic Crystals. Adv. Funct. Mater. 2010, 20, 1910– 1916.
- 254. López-García,M.; Galisteo-López, J.F.; Blanco, A.; Sanchez-Márcos, J.; Lopez, C.; García-Martín, A. Enhancement and directionality of spontaneous emission in hybrid self-assembled photonic-plasmonic crystals. Small 2010, 6, 1757–1761.
- 255. Wang, W.; Asher, S.A. Photochemical Incorporation of Silver Quantum Dots in Monodisperse Silica Colloids for Photonic Crystal Applications. J. Am. Chem. Soc. 2001, 123, 12528–12535.
- 256. Cai, Z.; Liu, Y.J.; Leong, E.S.P.; Teng, J.; Lu, X. Highly ordered and gap controllable two-dimensional nonclose-packed colloidal crystals and plasmonicphotonic crystals with enhanced optical transmission. J. Mater. Chem. 2012, 22, 24668–24675.

- 257. Ding, B.; Bardosova, M.; Pemble, M.E.; Korovin, A.V.; Peschel, U.; Romanov, S.G. Broadband omnidirectional diversion of light in hybrid plasmonic-photonic heterocrystals. Adv. Funct. Mater. 2011, 21, 4182–4192.
- 258. Vasquez, Y.; Kolle, M.; Mishchenko, L.; Hatton, B.D.; Aizenberg, J. Three-Phase Co-assembly: In Situ Incorporation of Nanoparticles into Tuneable, Highly Ordered, Porous Silica Films. ACS Photonics 2014, 1, 53–60
- 259. Pantalei, S.; Zampetti, E.; Macagnano, A.; Bearzotti, A.; Venditti, I.; Russo, M.V. Enhanced sensory properties of a multichannel quartz crystal microbalance coated with polymeric nanobeads. Sensors 2007, 7, 2920–2928.
- 260. Bearzotti, A.; Macagnano, A.; Pantalei, S.; Zampetti, E.; Venditti, I.; Fratoddi, I.; Russo, M.V. Alcohol vapors sensory properties of nanostructured conjugated polymer. J. Phys. Condens. Matter 2008, 20, 474207.
- 261. Cushing, S.K.; Hornak, L.A.; Lankford, J.; Liu, Y.; Wu, N. Origin of localized surface plasmon resonances in thin silver film over nanosphere patterns. Appl. Phys. A 2011, 103, 955–958.
- 262. Porcaro, F.; Carlini, L.; Ugolini, A.; Visaggio, D.; Luisetto, I.; Visca, P.; Fratoddi, I.; Venditti, I.; Simonelli, L.; Marini, C.; et al. Synthesis and Structural Characterization of Silver Nanoparticles Stabilized with 3-Mercapto-1-Propansulfonate and 1-Thioglucose Mixed Thiols for Antibacterial Applications. Materials 2016, 9, 1028.
- 263. Prosposito, P.; Mochi, F.; Ciotta, E.; Casalboni, M.; Venditti, I.; Fontana, L.; Testa, G.; Fratoddi, I. Hydrophilic silver nanoparticles with tuneable optical properties: Application for the detection of heavy metals in water. Beilstein J. Nanotechnol. 2016, 7, 1654–1661.
- 264. Xu, M.-F.; Zhu, X.-Z.; Shi, X.-B.; Liang, J.; Jin, Y.; Wang, Z.-K.; Liao, L.-S. Plasmon resonance enhanced optical absorption in inverted polymer/fullerene solar cells with metal nanoparticle-doped solutionprocessable TiO2 layer. ACS Appl. Mater. Interfaces 2014, 5, 2935–2942.
- 265. Cai, Z.; Leong, E.S.P.; Wang, Z.; Niu, W.; Zhang, W.; Ravaine, S.; Yakovlev, N.L.; Liu, Y.J.; Teng, J.; Lu, X. Sandwich-structured Fe2O3@SiO2@Au nanoparticles with magnetoplasmonic responses. J. Mater. Chem. C 2015, 3, 11645–11652.
- 266. Yang, Q.; Zhu, S.; Peng, W.; Yin, C.; Wang, W.; Gu, J.; Zhang, W.; Ma, J.; Deng, T.; Feng, C. Bioinspired fabrication of hierarchically structured, pH-tuneable photonic crystals with unique transition. ACS Nano 2013, 7, 4911–4918.
- 267. Gu, Z.-Z.;Horie, R.; Kubo, S.; Yamada, Y.; Fujishima,A.; Sato,O. Fabrication of aMetal-Coated Three-DimensionallyOrdered Macroporous Film and its Application as a Refractive Index Sensor. Angew. Chem. Int. Ed. 2002, 41, 1153–1156.