

Probiotic *Lactobacillus sp.* Improved Performance of Broiler Chicken: A Review

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ABSTRACT

Probiotic *Lactobacillus sp.* is considered a potential resource for improving the performance of broiler chicken in Indonesia. Administration of *Lactobacillus sp.* Probiotics could increase the Average Daily Gain (ADG) and body weight of broiler chickens by increasing intestinal villi permeability, so that nutrient absorption could be optimal. Use of *Lactobacillus sp.* in the ration is often combined with other probiotics, prebiotics, phytoadditives, and organic acids to support the feed intake and feed conversion ratio. *Lactobacillus sp.* also able to produce various bioactive compounds such as the enzyme manase which could be in symbiosis with *mannan-oligosaccharides* (MOS) to increase the bodyweight of broiler chickens and bacteriocins that against pathogenic bacteria such as *Salmonella typhimurium*, *Staphylococcus aureus* and *Bacillus cereus*. Probiotic metabolic activity of *Lactobacillus sp.* could suppress the growth of pathogenic bacteria by creating an acid digestive environment. In addition, immunity and hematological conditions such as erythrocytes, hematocrit, leukocytes, MCV, MCH, and MCHC were maintained well by *Lactobacillus sp.* supplementation in the ration. This review aims to highlight the potential of probiotic *Lactobacillus sp* to modulate broiler microbiome and metabolic status to enhance performance and health.

Keywords: Broiler chicken, *Lactobacillus sp.*, Production performance, Probiotics

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INTRODUCTION

Antibiotics have been used as ingredients in animal feed mixtures in Indonesia since 1946 with the aim of improving digestibility, growth, and health of livestock. However, its use as a growth driver has the potential to stimulate resistance (Krisnan *et al.*, 2019). Therefore, the Indonesian government has issued a regulation concerning the prohibition of the administration of antibiotic growth promoters in Law no. 18/2009 juncto No.41 / 2014 concerning Animal Husbandry and Animal Health which states the prohibition of the administration of feed mixed with certain hormones and/or feed additive antibiotics with the classification of veterinary drugs listed in the Minister of Agriculture Regulation No.14 / Permentan / PK .350/5/2017. The prohibition of antibiotics in feed is also strengthened by the issuance of MOA No. 22/2017 concerning Registration and Distribution of Feed, which requires a statement not to use antibiotic growth promoters in feed formulas produced by producers. In the previous study by Cardinal *et al.*, (2019) found that withdrawing antibiotic growth promoters from the broiler diet increased production costs, which in turn increased the price of poultry meat. Even though the consumption of chicken meat in Indonesia is about 6.1 kg per capita on average, this value is still lower than in other countries. Therefore, efforts are needed to replace the role of antibiotics growth promoters with an affordable, safe, and natural additive in the feed.

Currently, alternatives used for antibiotics include probiotics, prebiotics, competitive exclusion, enzymes,

and organic acids. The administration of probiotics is expected to be a candidate for AGP substitution. Probiotics are non-pathogenic living organisms present in some foods which have positive effects on the health of the hosts if they enter the body in sufficient amounts (Mousavi *et al.*, 2018). The most selected probiotics are gram-positive bacteria belonging to *Bacillus*, *Enterococcus*, *Lactobacillus*, *Pediococcus*, and *Streptococcus*. It is not only bacteria that are used as probiotics; yeast and fungi have been used, such as *Saccharomyces cerevisiae* and *Kluyveromyces* (Selaledi *et al.*, 2020).

The presence of *Lactobacillus sp.*, a common microbiota in the chicken broiler gastrointestinal tract, is essential to maintaining the ecological balance in the microbiota. The supplementation of probiotics to feed could maintain the balance of microflora in the digestive tract and inhibit pathogenic bacteria, increase digestive enzyme activity, reduce ammonia production, increase feed intake and digestion, neutralize enterotoxins and stimulate the immune system. Isolate *Lactobacillus sp.* could be taken from native Indonesian chicken which has great potential (Yulianto and Lokapirnasari, 2018).

Probiotic Classification

Probiotics are classified by observing the similarities of qualities, such as physical characteristics, metabolic needs, and metabolic end products. These similarities grouped into one genus, while the special characteristics that exist in one individual and do not exist in another individual will become the name of the species. The general classification of probiotics could be seen in table 1.

Table 1. The general classification of probiotics

Genus	Species	e.g., Strain	Ref.
Lactobacillus	murinus	Ar3	Harimurti <i>et al.</i> , 2010
		CR147	Harimurti <i>et al.</i> , 2010
	plantarum	BS22	Zeng <i>et al.</i> , 2018
		B1	Peng <i>et al.</i> , 2016
	acidophilus	D2	Cesare <i>et al.</i> , 2020
		A3B3	Sa'diyah <i>et al.</i> , 2020

	casei	CECT 4043	Sethiya NK, 2016
		Shirota	Sethiya NK, 2016
	salivarius	I72	Damayanti <i>et al.</i> , 2012
	reuteri	C1	Mookiah <i>et al.</i> , 2014
		C10	Mookiah <i>et al.</i> , 2014
		C16	Mookiah <i>et al.</i> , 2014
	agilis	JCM 1048	Lan <i>et al.</i> , 2004
		LA73	Ren <i>et al.</i> , 2019
	paracasei	L1	Xu <i>et al.</i> , 2019
	fermentum	CCM 7514	S'efcova' <i>et al.</i> , 2021
rhamnosus	TB1	Bouzaine <i>et al.</i> , 2005	
	JYLR-005	Liu <i>et al.</i> , 2021	
Streptococcus	thermophilus	Kp2	Harimurti and Ariyadi , 2014
		Kd2	Harimurti and Ariyadi , 2014
Pediococcus	acidilactici	Kd6	Harimurti <i>et al.</i> , 2010,
		DSM 20284	Damayanti <i>et al.</i> , 2014
	pentosaceus	Db9	Konosonoka <i>et al.</i> , 2015
		TMU457	Konosonoka <i>et al.</i> , 2015
		NP6	Khochamit <i>et al.</i> , 2020
Bacillus	subtilis	DSM 32315	Whelan <i>et al.</i> , 2019
		29784	Jacquier <i>et al.</i> , 2019
		PB6	Jayaraman <i>et al.</i> , 2017
		DSM17299	Knap <i>et al.</i> , 2011
		B21	Musa <i>et al.</i> , 2019
Saccharomyces	cerevisiae	CECT 1891	Pizzolitto <i>et al.</i> , 2013
	bouardii	CNCM I-1079	Massacci <i>et al.</i> , 2019
Aspergillus	oryzae	GB-107	Hong <i>et al.</i> , 2004
		3.042	Feng <i>et al.</i> , 2007

Among the various kinds of microorganisms used as probiotics, lactic acid bacteria (LAB) are the type of microorganism that is most widely used. Lactic acid bacteria are bacteria that could produce lactic acid as a result of the fermentation of carbohydrates. Lactic acid works to prevent the development of pathogenic microorganism; therefore, it has the potential as probiotics (Purwandhani and Rahayu, 2003). The dominant lactic acid microbiota in broiler GIT is *Lactobacillus reuteri*, *L. salivarius*, *L. agilis*, and *L. acidophilus*, about 10.19×10^9 CFU/ml in the small intestine and 10^{10} to 10^{11} cell / g in the large intestine (Manin *et al.*, 2010).

Effect Probiotic *Lactobacillus sp.* on ADG and Final Body Weight

Wahyudi and Hendraningsih (2007) showed that lactic acid *Lactobacillus casei* could increase intestinal villi permeability, nutrient adsorption, and digestibility, maintain the health of broiler chickens and resulted enzymes capable of digesting fiber, fat, and protein. Therefore, the nutrients absorbed properly to support the muscle development of broilers. These benefit mechanisms directly affect average body weight and feed conversion (Sari and Akbar, 2019; Pertiwi *et al.*, 2019). In similar research, the combination of 18% protein with 1.2% dahlia tuber inulin and 1.2 ml of *Lactobacillus sp.* was able to result intestinal villi height and improve protein digestibility (Purbarani *et al.*, 2019). By increasing the height of the intestinal villi, the absorption of feed nutrients could be optimized well.

Lactobacillus sp. found in the natural digestive system of chickens produced the enzyme manase which selectively

bind to *manna-oligosaccharides* (Selaledi, *et al.*, 2020). Administration of *mannan-oligosaccharides* (MOS) in broilers could increase daily body weight by 4-8% (Sinovec *et al.*, 2005). Various bioactive compounds produced by probiotic *Lactobacillus sp.* could be seen in table 2. In Contrast, Aurora *et al.*, (2020) reported that administration prebiotic *fructooligosaccharide* (FOS) with a combination of *Lactobacillus achidophilus* as much as 2 ml/kg of the ration could increase bone strength and length but has not increased weight gain to chicken bones. The tibia and femur bone weight did not increase significantly, presumably due to there has been a change in the structure of the bone constituent matrix due to increased absorption of Ca and protein, but it has not yet affected bone weight. *Lactobacillus sp.* could be combined with other beneficial microorganisms as probiotics. Mahajan *et al.*, (2000) reported a significant increase in the weight of viscera, fresh carcass weight, cold storage carcass weight, and carcass percentage of broilers fed with probiotics combination of *Lactobacillus sp.* and *Saccharomyces sp.*

The administration of multistrain probiotics between *Lactobacillus murinus*, *Streptococcus thermophilus*, and *Pediococcus acidilactici* improved small intestine morphology and integrity in broiler chickens by the develop villi height and depth of intestinal crypt (Harimurti and Sutriswati, 2009). Supplementation of 10^8 CFU/ml/bird daily of these types of probiotics could give a significant effect of weight gain increasing within 28 days (Harimurti *et al.*, 2010).

Table 2. Various bioactive compounds produced by probiotic *Lactobacillus sp.*

Bioactivities	Functions	References
Malic acid	Against <i>Escherichia coli</i> , <i>Salmonella typhimurium</i> , <i>Staphylococcus aureus</i> and <i>Bacillus cereus</i>	Baserisalehi <i>et al.</i> , 2017
Lactic acid		
Acetic acid		
Phenyllactic acid	Control foodborne pathogens, spoilage organisms, and contaminating yeasts	Kantachote <i>et al.</i> , 2010
Nisin	Forms pores in the cytoplasmic membrane in pathogenic bacteria	Gabriela <i>et al.</i> , 2016
Lactocin	Antimicrobial effect against <i>Escherichia coli</i> , <i>Salmonella typhimurium</i> , <i>Staphylococcus aureus</i> , and <i>Bacillus cereus</i>	Majid <i>et al.</i> , 2017
Reutericyclin	Antimicrobial effect	Mu <i>et al.</i> , 2018
Manase	Selectively bind manna-oligosaccharides	Selaledi <i>et al.</i> , 2020
Galactose	Enhanced their anti-inflammatory effects on macrophages	Chen <i>et al.</i> , 2019
Lipoteichoic acids	Reduces leaky gut and inflammation	Wang <i>et al.</i> , 2019
Serine	Anti-inflammatory response in dendritic cells from IBD patients	Al-Hassi <i>et al.</i> , 2014
Threonine		

Effect Probiotic *Lactobacillus sp.* on Feed Intake and Feed Conversion Ratio

Probiotics supplementation in feeds could maintain microflora balance in the digestive tract and inhibit pathogenic bacteria, increase digestive enzyme activity, decrease ammonia production, improve feed intake and digestion, neutralize enterotoxins and optimize the immune system (Manin *et al.*, 2010). Zhui *et al.*, (2018) showed that administration of probiotic *Lactobacillus acidophilus* in the ration, increased body weight, decreased FCR, and reduced mortality. In order to increase the effectiveness of prebiotics and probiotics in improving the productivity of broilers, prebiotics and probiotics are often used together. The combination of prebiotics and probiotics is known as a symbiotic. Prebiotics are a source of indigestible carbohydrates that affect the host by selectively stimulating the growth of some bacteria in the colon. (Selaledi *et al.*, 2020). Nutrients that are proposed to have prebiotic potential include fructans, oligofructose, inulin, fructooligosaccharides, galactan, galactooligosaccharides, resistant starch, pectin, fiber components, and milk oligosaccharides (Bird *et al.*, 2010). The content of these nutrients is found in plant extracts that are widely cultivated in Indonesia such as garlic (*Alium sativum*). Experiments conducted by Sunu *et al.*, (2020) showed that using 4 ml of garlic extract with the supplementation of *Lactobacillus acidophilus* at a ratio of 100: 1 ml was able to produce a feed consumption of 2908.92 ± 1.24 g and feed conversion of 1.24 ± 0.04 g within 35 days.

Naik *et al.*, (2000) evaluated the effect of different probiotics between *Lactobacillus acidophilus*, *Saccharomyces cerevisiae*, and their combination) on the performance of broilers and reported that supplementation of *Lactobacillus* to the basal diet at 0.05% improved feed efficiency in broilers as compared to the control group. The combination of several probiotics *Lactobacillus plantarum*, *Lactobacillus bulgaricus*, *Lactobacillus acidophilus*, *Lactobacillus rhamnosus*, *Bifidobacterium bifidum*, *Streptococcus thermophilus*, *Enterococcus faecium*, and *Bacillus subtilis* (10^9) yields a good feed conversion on broiler (Rocha *et al.*, 2010). Harimurti *et al.*, (2010) showed the administration of *Lactobacillus murinus*, *Streptococcus thermophilus*, and *Pediococcus acidilactici* about 108 CFU /ml/bird daily provides a feed intake of 1359.72 g/bird with a feed conversion rate of 1.93 within 28 days.

Experiments conducted by Pasi *et al.*, (2018) for six weeks resulted in a feed intake of 3285.87 ± 7.39 g and FCR 2.28 ± 0.04 using a combination probiotic between *Lactobacillus sporogenes* + live yeast 0.02% + 0.2% g /chick/pen. The administration of *Lactobacillus* could also be combined with organic acids. Treatment of organic acids and probiotics leads to better performance in the examined properties in male Ross 308 broilers compared to the feed containing antibiotics especially for feed intake, weight gain, FCR, and abdominal fat percentage (Beheshti *et al.*, 2009).

Effect Probiotic *Lactobacillus sp.* on Animal Health

Normal flora microorganisms in the digestive tract will compete with each other for nutrition and attachment sites in the gastrointestinal ecosystem. Pathogenic bacteria could initiate infection disease if they have dominated the population of the total microorganisms in the digestive tract by suppress beneficial bacteria activity, increase pH and release a toxin (Sunu *et al.*, 2020). The immune system of broilers in the early stage has not developed well and susceptible to bacterial infections caused by *Campylobacter jejuni*, *C. perfringens*, *Salmonella enterica*, and *Escherichia coli* (Ohimain and Ofongo, 2012). Therefore, it suggested intestinal floral microbes balance need to be maintained to support the health of chickens to start growing fast in the starter phase.

Lactic acid bacteria as probiotics in the digestive tract could inhibit the activity and growth of pathogenic microbes through the production of organic acids, hydrogen peroxide (H_2O_2), and bacteriocins (Taheri *et al.*, 2009). A similar study by Sari and Akbar (2019) found that lactic acid bacteria not only producing bacteriocin but also resulted physiological effects on broiler health, as a feed supplement, drugs (such as natural antibiotics), therapeutic effects (such as hypo cholesterol and antihypertensive, prevention of diarrhea). In addition, some types of lactic acid bacteria could lower the pH level of the digestive tract, which disturb pathogenic bacteria metabolism due to unsuitable environment condition for them (Mousavi *et al.*, 2018). The inhibition mechanism of bacteriocin occurred in two phases. The first phase was bacteriocin absorption of specific and nonspecific receptors on target bacterial membrane cells. During this phase, the bacteriocins became sensitive, especially to proteolytic enzymes. A second phase was irreversible and involves lethal changes in the sensitive strains (Damayanti *et al.*, 2012).

The administration of *Lactobacillus sp.* probiotics could restore the amount of microflora in the digestive tract and suppress the coliform population (Hamid et al., 2014). *L. reuteri*, *L. salivarius*, or *Lactobacillus sp.* treatment could inhibit the pathogen bacterial such as *Enterococcus faecalis*, *Enterococcus faecium*, *Listeria monocytogenes*, and *Salmonella sp.* (Lima et al., 2007). Meanwhile, *L. fermentum* and *L. rhamnosus* were able to fight against *E. coli* and *S. pullorum* (Torshizi et al., 2008).

In the similar research by Olnood et al. (2015), reported that the administration of multi strain *Lactobacillus spp.* probiotic in broilers diet elevated the number of total Enterobacteria and anaerobic bacteria in small intestine especially in the ileum and caeca, and the amount of lactic acid bacteria in the caeca, due to the ability to co- and auto aggregate. Commonly, microorganisms demonstrating a high auto-aggregation potential result a good adhesion to the mucus (Jha et al., 2020). One of the indications of immunity organ health improvement of broilers was indicated by the increase of bursa fabricius weight as the central of lymphoid organ (Purbarani et al., 2019). The size of the bursa of fabricius correlates with the high number of B cells matured and produced by the chicken. Zhang et al., (2012) reported that administration of the probiotic *Lactobacillus paracasei* resulted higher percentage of lymphoid organs (Fabricius bursa and spleen) on the poultry to support their health and production performance.

A similar study by Hidayat, (2020) showed that supplementation *Lactobacillus paracasei* about of 5 ml/day increased the weight of immune organs such as 0.194% in the bursa of Fabricius, 0.20% in the spleen, and 0.32% in the thymus of the total weight of the lymphoid organs, It also improves hematological level significantly as health biomarker of broilers, include erythrocyte (2.67 x 10⁶ mm³), hematocrit (29.2%), leukocytes (21.1 x 10³ / mm³), MCV (109.82 - 111.53 fl), MCH (32.77 - 34.11 Pg), and MCHC (29.96% - 31.14%).

Lactobacillus sp. which is used as a fish meal fermentation as broilers feed could increase serum lipid performance in broiler chickens. Sumarsih et al., (2010) showed that the administration of the fermented fish meal with *Lactobacillus sp.* as much as 4 - 8% in the ration reduced the average LDL cholesterol (66.86) and triglycerides (82.60). Cholesterol status is highly related to health conditions. There are two types of cholesterol, which give a different impact on health, is LDL cholesterol provides negative effects, while HDL cholesterol has a positive impact on health, while triglycerides whose density is reduced will turn into LDL (Sumarsih et al., 2010).

CONCLUSION

The administration of *Lactobacillus sp.* As probiotics have a beneficial effect on production performance and Animal Health. The application of *Lactobacillus sp.* could be an alternative of natural growth promoters to replace AGP in the broiler chicken's feed.

REFERENCES

- Krisnan R, Retnani Y, Tangendjaja B et al. In ovo Feeding of Butyric Acid Replacing Antibiotics Function to Increase Poultry Productivity. *Wartazoa*. 2019;29(1): 35-42
- Cardinal K., Kipper M., Andretta, I., et al. Withdrawal of antibiotic growth promoters from broiler diets: Performance indexes and economic impact. *Poult. Sci*. 2019;98: 6659-6667
- Mousavi, Hosseini HM, and Mirhosseini SA. A Review of Dietary Probiotics in Poultry. *J Appl Biotechnol Rep*. 2018;5(2): 48-54
- Selaledi LA, Hassan ZM, Manyelo TG, et al. Review the current status of the alternative use to antibiotics in poultry production: an african perspective. *Antibiotics*. 2020;9: 1-18
- Yulianto B and Lokapirnasari WP. Isolation and identification of lactic acid bacteria from the digestive tract of kampung chicken. *Philipp. J. Vet. Med*. 2018;55: 67-72
- Harimurti S, Nasroedin, Rahayu ES, and Kurniasih. Effect of indigenouse lactic acid bacteria probiotics on broiler performance. The 5th International Seminar on Tropical Animal Production Community Empowerment and Tropical Animal Industry. 2010: 391-394
- Zeng Y, Zeng D, Zhang Y, et al. *Lactobacillus plantarum* BS22 promotes gut microbial homeostasis in broiler chickens exposed to aflatoxin B₁. *J Anim Physiol Anim Nutr (Berl)*. 2018;102(1): 449-459
- Peng Q, Zeng XF, Zhu JL, et al. Effects of dietary *Lactobacillus plantarum* B1 on growth performance, intestinal microbiota, and short chain fatty acid profiles in broiler chickens. *Poult Sci*. 2016;95(4): 893-900
- Cesare AD, Sala C, Castellani G, et al. Effect of *Lactobacillus acidophilus* D2/CSL (CECT 4529) supplementation in drinking water on chicken crop and caeca microbiome. *PLoS One*. 2020; 15(1): 1-18
- Sa'diyah SN, Sukanto B, Wahyono F, et al. Penambahan Kominasi Ekstrak Buah Noni (*Morinda citrifolia L.*) dan *Lactobacillus acidophilus* dalam Pakan Terhadap Profil Lemak Darah Ayam Pedaging. *Jurnal Nutrisi Ternak Tropis*. 2020;3: 81-89
- Sethiya NK. Review on Natural Growth Promoters Available for Improving Gut Health of Poultry: An Alternative to Antibiotic Growth Promoters. *Asian Journal of Poultry Science*. 2016; 10 (1): 1-29
- Damayanti E, Herdian H, Angwar M, et al. Lactic acid bacterial screening from gastrointestinal digestive tract of native and broiler chicken for probiotic candidate purposes. *J.Indonesian Trop.Anim.Agric*. 2012;37(3): 168-175
- Mookiah S, Sieo CC, Ramasamy K, Abdullah N, et al. Effects of dietary prebiotics, probiotic and synbiotics on performance, caecal bacterial populations and caecal fermentation concentrations of broiler chickens. *J. Sci. Food Agric*. 2014;94: 341-348
- Lan PTN, Sakamoto M, and Benno Y. Effects of Two Probiotic *Lactobacillus* Strains on Jejunal and Cecal Microbiota of Broiler Chicken under Acute Heat Stress Condition as Revealed by Molecular Analysis of 16S rRNA Genes. *Microbiol. Immunol*. 2014;48 (12): 917-929
- Ren H, Vahjen H, Dadi T, et al. Synergistic Effects of Probiotics and Phytobiotics on the Intestinal Microbiota in Young Broiler Chicken. *Microorganisms*. 2019;7 (12): 1-23
- Feng J, Liu X, Xu ZR, et al. Effects of *Aspergillus oryzae* 3.042 fermented soybean meal on growth performance and plasma biochemical parameters in broilers. *Animal Feed Science and Technology*. 2007;134: 235-242
- S'efcova' M, Marco LA, Ce'sar LA, et al. The Probiotic *Lactobacillus fermentum* Biocenol CCM 7514 Moderates *Campylobacter jejuni*-Induced Body Weight Impairment by Improving Gut Morphometry

- and Regulating Cecal Cytokine Abundance in Broiler Chickens. *Animals*. 2021;11(1): 235
18. Bouzaine T, Dauphin RD, Thonart P, et al. Adherence and colonization properties of *Lactobacillus rhamnosus* TB1, a broiler chicken isolate. *Microbiology* 2005;40: 391-396
 19. Liu, F., Kong, A., Fu, P. et al. *Lactobacillus rhamnosus* JYLR-005 Prevents Thiram-Induced Tibial Dyschondroplasia by Enhancing Bone-Related Growth Performance in Chickens. *Probiotics & Antimicro. Prot.* 2021;13: 19-31
 20. Harimurti S and Ariyadi B. Effect of Lactic Acid Bacteria on the Intestinal Structure and the Expression of Ileal Tight Junction Molecule Claudins in Broiler. Proceedings of the 16th AAAP Animal Science Congress. 2014;2: 1210-1213
 21. Damayanti E, Julendra H, Sofyan A, et al. Bile Salt and Acid Tolerant of Lactic Acid Bacteria Isolated from Proventriculus of Broiler Chicken. *Media Peternakan*. 2014;37(2): 80-86
 22. Konosonoka IH, Osmane B, Cerina B, et al. Feeding Technology Impact on Broiler Productivity and Intestinal Tract Microflora. *Jelgava*. 2015;5: 20-22
 23. Khochamit N, Siripornadulsil S, Sukon P, et al. *Bacillus subtilis* and Lactic Acid Bacteria Improve the Growth Performance and Blood Parameters and Reduce *Salmonella* infection in broilers. *Veterinary World*. 2020;13(12): 2663-2672
 24. Whelan RA, Doranalli K, Rinttilä T, et al. The impact of *Bacillus subtilis* DSM 32315 on the pathology, performance, and intestinal microbiome of broiler chickens in a necrotic enteritis challenge. *Poult Sci*. 2019;98(9): 3450-3463
 25. Jacquier V, Nelson A, Jilali M, et al. *Bacillus subtilis* 29784 induces a shift in broiler gut microbiome toward butyrate-producing bacteria and improves intestinal histomorphology and animal performance. *Poult Sci*. 2019;98(6): 2548-2554
 26. Jayaraman S, Das PP, Saini PC, et al. Use of *Bacillus Subtilis* PB6 as a potential antibiotic growth promoter replacement in improving performance of broiler birds. *Poult Sci*. 2017;96(8): 2614-2622
 27. Knap I, Kehlet AB, Bennedsen M, et al. *Bacillus subtilis* (DSM17299) significantly reduces *Salmonella* in broilers. *Poult Sci*. 2011;90(8): 1690-1694
 28. Musa BB, Duan Y, Khawar H, et al. *Bacillus subtilis* B21 and *Bacillus licheniformis* B26 improve intestinal health and performance of broiler chickens with *Clostridium perfringens*-induced necrotic enteritis. *J Anim Physiol Anim Nutr (Berl)*. 2019;103(4): 1039-1049
 29. Pizzolitto RP, Armando MR, Salvano MA, et al. Evaluation of *Saccharomyces cerevisiae* as an anti-aflatoxicogenic agent in broiler feedstuffs. *Poultry Science*. 2013;92 (6): 1655-1663
 30. Massacci FR, Lovito C, Tofani S, et al. Dietary *Saccharomyces cerevisiae boulardii* CNCM I-1079 Positively Affects Performance and Intestinal Ecosystem in Broilers during a *Campylobacter jejuni* Infection. *Microorganisms*. 2019;7(12): 596
 31. Hong KL, Lee CH, and Kim SW. *Aspergillus oryzae* GB-107 fermentation improves nutritional quality of food soybeans and feed soybean meals. *Journal of Medicinal Food*. 2004;7: 430-435
 32. Purwandhani, SN and Rahayu, ES. Isolation and selection of the lactobacilli which are considered probiotic agents. *Agritech*. 2003;23(2): 67-74
 33. Manin F, Hendalia E, and Aziz A. Isolation and production of lactic acid bacteria and *Bacillus* sp. from gastrointestinal non ras chicken origin Gambut land as source of probiotic. *Scientific Journal of Animal Science*. 2010;8(5): 221-228
 34. Wahyudi A. and L.Hendraningsih. 2007. Probiotik, Konsep dan Penerapan pada Ternak ruminansia. UMM Press. Malang
 35. Sari RM and Akbar SA. The influence of lactic acid bacteria *Lactobasillus casei* for performance of broiler. *IOP Conf. Series: Earth and Environmental Science*. 2019;347: 1-4
 36. Pertiwi H, Sidik R, Sabdoningrum EK, Dadi TB. 2019. Carcass Quality of Broiler Supplemented with *Spirulina*, Kelor Leaves (*Moringa oleifera*), and Probiotic. *Indian Vet. J.*, 96 (11) : 39 - 41
 37. Purbarani SA, Wahyuni HI, and Suthama N. Dahlia inulin and *Lactobacillus* sp. in step down protein diet on villi development and growth of KUB chickens. *Tropical Animal Science Journal*. 2019;42(1): 19-24
 38. Sinovec Z and Markovic R. Use of pre-biotics in poultry nutrition. *Biotechnol*. 2005;21: 235-239
 39. Aurora NE, Mahfudz LD, and Sarjana TA. The potential of garlic and *Lactobacillus acidophilus* as synbiotics on the characteristics of broiler chicken bones. *Jurnal Sain Peternakan Indonesia*. 2020;15 (4): 375-382
 40. Mahajan P, Sahoo J, and Panda PC. Effect of probiotic (Lacto-Sacc) feeding, packaging methods and seasons on the microbial and organoleptic qualities of chicken meat balls during refrigerated storage. *J Food Sci Technol (Mysore)*. 2000;37(1): 67-71
 41. Harimurti S and Rahayu ES. Study of Competitive Exclusion of *Salmonella Pullorum* by Probiotic Strains in The Broiler Chickens. Oral presentation on the 5th Asian Conference on Lactic Acid Bacteria : Microbes in Disease Prevention & Treatment. Singapore, 1st - 3rd July. 2009;41(2): 134-141
 42. Harimurti S, Nasroedin, Rahayu ES, and Kurniasih. Effect of indigenous lactic acid bacteria probiotics on broiler performance. The 5th International Seminar on Tropical Animal Production Community Empowerment and Tropical Animal Industry. 2010: 391-394
 43. Baserisalehi M, Jamali F, Moradi P, et al. Chemical Analysis of Bioactive Compound Produced by *Lactobacillus reuteri* Isolated from Domestic Chickens in Iran. *Biomed J Sci & Tech Res*. 2017; 1(7): 1964-1966
 44. Kantachote D, Prachyakij P, Charernjiratrakul W et al. Characterization of the antiyeast compound and probiotic properties of a starter *Lactobacillus plantarum* DW3 for possible use in fermented plant beverages. In: *Electronic Journal of Biotechnology*. 2010;13: 5
 45. Gabriela N. Tenea and Lucia Yépez. Bioactive Compounds of Lactic Acid Bacteria. Case Study: Evaluation of Antimicrobial Activity of Bacteriocin-producing *Lactobacilli* Isolated from Native Ecological Niches of Ecuador. *Intech*. 2016;8: 149-167
 46. Majid B, Fatemeh J, Parviz M et al. Chemical Analysis of Bioactive Compound Produced by *Lactobacillus reuteri* Isolated from Domestic Chickens in Iran. *Biomed J Sci & Tech Res*. 2017;1(7): 1964-1966
 47. Mu Q, Tavella VJ, and Luo XM. Role of *Lactobacillus reuteri* in human health and diseases. *Front. Microbiol*. 2018;9: 757

48. Chen, YC, Wu YJ, Hu CY. Monosaccharide composition influence and immunomodulatory effects of probiotic exopolysaccharides. *Int. J. Biol. Macromol.* 2019;133: 575–582
49. Wang S, Ahmadi S, Nagpal R, et al. Lipoteichoic acid from the cell wall of a heat killed *Lactobacillus paracasei* D3-5 ameliorates aging-related leaky gut, inflammation and improves physical and cognitive functions: From *C. elegans* to mice. *Geroscience.* 2019;42: 333–352
50. Al-Hassi HO, Mann ER, Sanchez B, et al. Altered human gut dendritic cell properties in ulcerative colitis are reversed by *Lactobacillus plantarum* extracellular encrypted peptide STp. *Mol. Nutr. Food Res.* 2014;58: 1132–1143
51. Zhui Li, Weiwei W, and, Yuming L. Effects of *Lactobacillus acidophilus* on the growth performance and intestinal health of broilers challenged with *Clostridium perfringens*. *J. Anim. Sci. Biotechnol.* 2018;9: 25–35
52. Bird AR, Conlon MA, Christophersen CT, and Topping DL. Resistant starch, large bowel fermentation and a broader perspective of prebiotics and probiotics. *Benef. Microbes.* 2010;1: 423–431
53. Rocha, AP, Abreu RD, Costa MC et al. Prebiotics, organic acids and probiotics in broiler feed. *Revista Brasileira de Saúde e Produção Animal.* 2010;11(3): 793-801
54. Pasi SK, Bais B, Dhaka CS, et al. Studies on the effect of *Lactobacillus* with and without supplementation of live yeast on growth performance of broilers in arid zone of Rajasthan. *International Journal of Chemical Studies.* 2018; 6(3): 434-437
55. Beheshti MMH, Rezaei M, Niknafs F, Sayyazadeh H. Effect of combined probiotic and organic acid on some blood parameters and immune system of broiler chicks. *Proceedings of the 2nd Mediterranean Summit of World Poultry Science Association.* 2009: 4-7
56. Sunu P, Sunarti D, Mahfudz LD et al., Effect of synbiotic from *Allium sativum* and *Lactobacillus acidophilus* on hematological indices, antioxidative status and intestinal ecology of broiler chicken, *Journal of the Saudi Society of Agricultural Sciences*, <https://doi.org/10.1016/j.jssas.2020.12.005>
57. Naik DG, Javedmulla A and Shivakumar MC (2000). Performance of broilers supplemented with probiotics. *Karnataka Journal of Agricultural Sciences*, 13(4): 957-960
58. Ohimain EI and Ofongo RTS. The effect of probiotic and prebiotic feed supplementation on chicken health and gut microflora: a review. *Int. J. Anim. Vet. Adv.* 2012;4(2): 135-143
59. Taheri HR, Moravej H, Tabandeh F, et al. Screening of lactic acid bacteria toward their selection as a source of chicken probiotic. *Poultry Science.* 2009;88(8): 1586-1593
60. Hamid IS, Rahardjo BPS, and Gabriela M. The potential of giving synbiotics at different ages on the histology of ileum in broilers. *J. Vet. Medika.* 2014;7: 114-119
61. Lima ET, Filho RLA, Okamoto AS, et al. Evaluation in vitro of the antagonistic substances produced by *Lactobacillus* spp. 2007;71: 103-107
62. Torshizi MAK, Rahimi S, Mojgani N. Screening of indigenous strains of lactic acid bacteria for development of a probiotic for poultry. *Asian-Aus. J. Anim. Sci.* 2008;21: 1495– 1500
63. Olnood CG, Beski SSM, Choct M, Iji PA. Novel probiotics: Their effects on growth performance, gut development, microbial community and activity of broiler chickens. *Anim. Nutr.* 2015, 1, 184–191.
64. Jha R, Das R, Oak S, Mishra P. Probiotics (Direct-Fed Microbials) in Poultry Nutrition and Their Effects on Nutrient Utilization, Growth and Laying Performance, and Gut Health: A Systematic Review. *Animals* 2020, 10, 1863; doi:10.3390/ani10101863
65. Zhang ZF, Zhou TX, Ao X, et al. Effects of β -glucan and *Bacillus subtilis* on growth performance, blood profiles, relative organ weight and meat quality in broilers fed maize–soybean meal-based diets *Livest. Sci.* 2012;150: 419–24
66. Hidayat MN, Malaka R, Agustina L. Effect of probiotic *Lactobacillus paracasei* on hematology and relative weight of lymphoid organs of broiler. *IOP Conf. Series: Earth and Environmental Science.* 2020; 492: 1-7
67. Sumarsih S, Yudiarti T, Utami CS, et al. The influence of using fish fermented by lactic acid bacteria as feed substitution on serum lipid profile of broilers. *J.Indonesian Trop.Anim.Agric.* 2010;35(2): 124-128